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New Taxa and Combinations in Onciderini Thomson, 1860 (Coleoptera:
Cerambycidae: Lamiinae) from Central and South America, with notes
on additional taxa

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New Taxa and Combinations in Onciderini Thomson, 1860 (Coleoptera: Cerambycidae: Lamiinae) from Central and South America, with notes on additional taxa

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Abstract. *Touroultia*, a **new genus** of Onciderini Thomson, 1860 (Coleoptera: Cerambycidae: Lamiinae) is described and illustrated. Five **new species** of Onciderini are also described and illustrated: *Jamesia ramirezi* from Costa Rica; *Peritrox marcelae* from French Guiana; *Touroultia swifti* from Ecuador; *Touroultia lordi* from French Guiana; *Trestoncideres santossilvai* from Brazil. Keys to the known species of *Peritrox* Bates, 1865; *Touroultia* gen. nov.; and *Trestoncideres* Martins and Galileo, 1990 are provided. The following **new synonymies** are proposed: *Calliphenges* Waterhouse, 1880 (Colobotheni) = *Malthonea* Thomson, 1864 (Desmiphorini); *Paraclitemnestra* Breuning, 1974 (Onciderini) = *Jamesia* Jekel, 1861 (Onciderini); *Orteguaza* Lane, 1958 (Apomecynini) = *Clavidesmus* Dillon and Dillon, 1946 (Onciderini). The following **new combinations** are proposed: *Clavidesmus funerarius* (Lane, 1958) (Onciderini); *Clavidesmus lichenigerus* (Lane, 1958) (Onciderini); *Ischiocentra insulata* (Rodrigues and Mermudes, 2011); *Malthonea cuprascens* (Waterhouse, 1880) (Desmiphorini); *Touroultia obscurella* (Bates, 1865) (Onciderini). The following species is **restored to original combination**: *Jamesia lineata* Fisher, 1926 (Onciderini). The following **13 new country records** are reported: *Ataxia hovorei* Lingafelter and Nearn, 2007 (Pteropliini) (Haiti); *Carterica soror* Belon, 1896 (Colobotheni) (Ecuador); *Colobothea lunulata* Lucas, 1859 (Colobotheni) (Colombia); *Curius punctatus* (Fisher, 1932) (Curiini) (Haiti); *Cyclopeplus lacordairei* Thomson, 1868 (Anisocerini) (Colombia); *Iarucanga mimica* (Bates, 1866) (Hemilophini) (Ecuador); *Pirangoclytus latithorax* (Martins and Galileo, 2008) (Clytini) (Costa Rica); *Porangonycha princeps* (Bates, 1872) (Hemilophini) (Colombia); *Trestonia lateapicata* Martins and Galileo, 2010 (Onciderini) (Brazil); *Tulcus dimidiatus* (Bates, 1865) (Onciderini) (Colombia); *Unaporanga cincta* Martins and Galileo, 2007 (Hemilophini) (Colombia); *Zeale dubia* Galileo and Martins, 1997 (Hemilophini) (Colombia); *Zonotylus interruptus* (Olivier, 1790) (Trachyderini) (Colombia).

Key words. Key; Neotropical; New distribution record; New genus; New species; New synonymy; Taxonomy.

Resumen. *Touroultia*, un **nuevo género** de Onciderini Thomson, 1860 (Coleoptera: Cerambycidae: Lamiinae) es descrito e ilustrado. Cinco **nuevas especies** de Onciderini son descritas e ilustradas: *Jamesia ramirezi* de Costa Rica; *Peritrox marcelae* de Guayana Francesa; *Touroultia swifti* de Ecuador; *Touroultia lordi* de Guayana Francesa; *Trestoncideres santossilvai* de Brasil. Claves de las especies de *Peritrox* Bates, 1865; *Touroultia* gen. nov.; y *Trestoncideres* Martins y Galileo, 1990; son incluidas. Las siguientes **nuevas sinonimias** se proponen: *Calliphenges* Waterhouse, 1880 (Colobotheni) = *Malthonea* Thomson, 1864 (Desmiphorini); *Paraclitemnestra* Breuning, 1974 (Onciderini) = *Jamesia* Jekel, 1861 (Onciderini); *Orteguaza* Lane, 1958 (Apomecynini) = *Clavidesmus* Dillon y Dillon, 1946 (Onciderini). Las siguientes **nuevas combinaciones** se proponen: *Clavidesmus funerarius* (Lane, 1958) (Onciderini); *Clavidesmus lichenigerus* (Lane, 1958) (Onciderini); *Ischiocentra insulata* (Rodrigues y Mermudes, 2011); *Malthonea cuprascens* (Waterhouse, 1880) (Desmiphorini); *Touroultia obscurella* (Bates, 1865) (Onciderini). La siguiente especie es **restaurada a la combinación original**: *Jamesia lineata* Fisher, 1926 (Onciderini). Los siguientes **13 nuevos registros** de país se reportan: *Ataxia hovorei* Lingafelter y Nearn, 2007 (Pteropliini) (Haiti); *Carterica soror* Belon, 1896 (Colobotheni) (Ecuador); *Colobothea lunulata* Lucas, 1859 (Colobotheni) (Colombia);

Curius punctatus (Fisher, 1932) (Curiini) (Haiti); *Cyclopeplus lacordairei* Thomson, 1868 (Anisocerini) (Colombia); *Iarucanga mimica* (Bates, 1866) (Hemilophini) (Ecuador); *Pirangoclytus latithorax* (Martins y Galileo, 2008) (Clytini) (Costa Rica); *Porangonycha princeps* (Bates, 1872) (Hemilophini) (Colombia); *Trestonia lateapicata* Martins y Galileo, 2010 (Onciderini) (Brasil); *Tulcus dimidiatus* (Bates, 1865) (Onciderini) (Colombia); *Unaporanga cincta* Martins y Galileo, 2007 (Hemilophini) (Colombia); *Zeale dubia* Galileo y Martins, 1997 (Hemilophini) (Colombia); *Zonotylus interruptus* (Olivier, 1790) (Trachyderini) (Colombia).

Palabras Claves. Clave; Nueva especie; Nueva sinonimia; Nuevo género; Nuevo registro de país; Región neotropical; Taxonomía.

Introduction

The tribe Onciderini Thomson, 1860 (Cerambycidae: Lamiinae) is widely distributed in the New World from North America to southern South America. Nearn and Swift (2011) provided a brief review of the taxonomic history of the tribe. Onciderini currently consists of approximately 468 described species in 79 genera. It is worth noting that over half (52) of the 79 genera are either monotypic or have only two species. A phylogenetic analysis of the tribe has not been conducted and its monophyly remains untested. A morphological study and cladistic analysis of the tribe is forthcoming (Nearn and Miller in preparation).

During the process of producing a Lucid key to the genera of Onciderini (Nearn et al. 2011), several new taxa, taxonomic problems, and distribution records came to light (see Nearn and Swift 2011). Here we add a new genus and five new species, propose three synonymies, five new combinations, and add 13 new country records in the subfamilies Cerambycinae and Lamiinae.

Materials

Specimens from the following collections were examined and the following codens are used throughout the paper:

ACMS	American Coleoptera Museum, San Antonio, Texas, USA
BMNH	The Natural History Museum, London, United Kingdom
CMNH	Carnegie Museum of Natural History, Pittsburgh, Pennsylvania, USA
CUIC	Cornell University Insect Collection, Ithaca, New York, USA
DFPC	Denis Faure Private Collection, Kourou, French Guiana
EFGC	Edmund F. Giesbert Collection (at FSCA), Gainesville, Florida, USA
ENPC	Eugenio H. Nearn Private Collection, Albuquerque, New Mexico, USA
FSCA	Florida State Collection of Arthropods, Gainesville, Florida, USA
INBC	Instituto Nacional de Biodiversidad, Santo Domingo de Heredia, Heredia, Costa Rica
ISNB	Institut royal des Sciences naturelles de Belgique, Brussels, Belgium
ISPC	Ian P. Swift Private Collection, Orange County, California, USA
JLGC	Jean-Louis Giuglaris Private Collection, Matoury, French Guiana
JTPC	Julien Touroult Private Collection, Soyaux, France
MCNZ	Museu de Ciências Naturais, Fundação Zoobotânica do Rio Grande do Sul, Porto Alegre, Brazil
MNCR	Departamento de Historia Natural, Museo Nacional de Costa Rica, San José, Costa Rica
MNHN	Muséum National d'Histoire Naturelle, Paris, France
MNRJ	Museu Nacional, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Brazil
MUSM	Museo de Historia Natural Universidad Nacional Mayor de San Marcos, Lima, Peru
MZSP	Museu de Zoologia, Universidade de São Paulo, São Paulo, Brazil
NHRS	Swedish Museum of Natural History, Stockholm, Sweden
NMBA	Naturhistorisches Museum Basel, Basel, Switzerland
PHDC	Pierre-Henri Dalens Private Collection, Rémire-Montjoly, French Guiana
RRCC	Rolando Ramírez Campos Private Collection, Veragua, Costa Rica
SMFD	Forschungsinstitut und Naturmuseum Senckenberg, Frankfurt-am-Main, Germany

USNM	National Museum of Natural History, Smithsonian Institution, Washington, District of Columbia, USA
ZMHB	Museum für Naturkunde der Humboldt-Universität, Berlin, Germany
ZMSC	Bavarian State Collection of Zoology, Munich, Germany
ZMUC	Zoological Museum University of Copenhagen, Copenhagen, Denmark

Observations of specimens were made using a Max Erb stereomicroscope with 10× eyepieces. Photographs were taken with Visionary Digital's Passport Storm imaging system fitted with a Canon EOS 40D. Label data are verbatim and placed in quotes. Classification and distributional data are based on Monné (2005a, 2005b, 2012) and Monné and Bezark (2011).

Taxonomy

Jamesia Jekel, 1861: 259 (Lamiinae: Onciderini)

Type-species. *Lamia globifera* Fabricius, 1801 (original designation).

The genus *Jamesia* currently contains nine described species. Nearn et al. (2011) provided color photographs of seven species of this genus, including four type specimens.

Jamesia ramirezi Nearn and Tavakilian, sp. nov.

(Figures 1a–d)

Description. Female. Length 25.0–27.0 mm (measured from vertex to elytral apices), width 10.5–11.0 mm (measured across humeri). Habitus as in Fig. 1a. General form elongate-ovate, moderate to large-sized. Integument ferruginous or dark brown, entire body (except antennae and legs) with tawny pubescence, mottled with dark brown maculae of various sizes; elytra with two large, dark brown, irregularly-shaped maculae near middle.

Head with frons elongate, about 3/4 times width of lower eye lobe (as in Fig. 1c). Eyes with lower lobes large, ovate-oblong. Genae subquadrate to trapezoidal, distinctly shorter than lower eye lobes. Antennae about 1.3 times longer than body; antennal tubercles prominent, narrowly separated, contiguous at base; tubercles armed at apex with small, blunt tubercle; scape robust, slightly bowed, gradually expanded to apex. Antennal formula based on antennomere III: scape=0.80; II=0.06; III=1; IV=0.81; V=0.63; VI=0.59; VII=0.53; VIII=0.45; IX=0.45; X=0.45; XI=0.45.

Pronotum roughly conical, distinctly wider at base, transverse, about 1.5 times as wide as long, sides feebly arcuate (Fig. 1d); disk at middle near base with a feebly elevated, median tubercle, either side of middle with three blunt tubercles arranged in a triangle; one apical and one transverse sulcus, and a more distinct oblique sulcus laterally which continues down the side.

Scutellum transverse, apex rounded.

Elytra about 1.75 times as long as width at humeri (Fig. 1a), about 4.5 times as long as pronotal length, about 1.6 times broader basally than pronotum at widest (at base); sides slightly sinuate, distinctly attenuate to apices, elytral apices individually rounded; base of each elytron somewhat gibbose, gibbositities each with prominent granules, rest of base with minute scattered granules, widely separated punctures placed one in each dark brown pubescent spot; humeri prominent, anterior margin oblique, the angle with a moderate-sized shining tubercle; sides below humeri with a few small granules.

Venter with procoxae large, globose, not uncate; narrowest area of prosternal process between procoxae about 1/4 as wide as procoxal cavity; apex of prosternal process subtriangular. Mesosternal process about as wide as mesocoxal cavity; mesosternal process deeply emarginate. Fifth sternite nearly 2 times as long as IV, with a median triangular impression.

Legs moderate in length; femora robust, gradually expanded to apex; tibiae slightly expanded apically; metafemora about 1/3 as long as elytra.

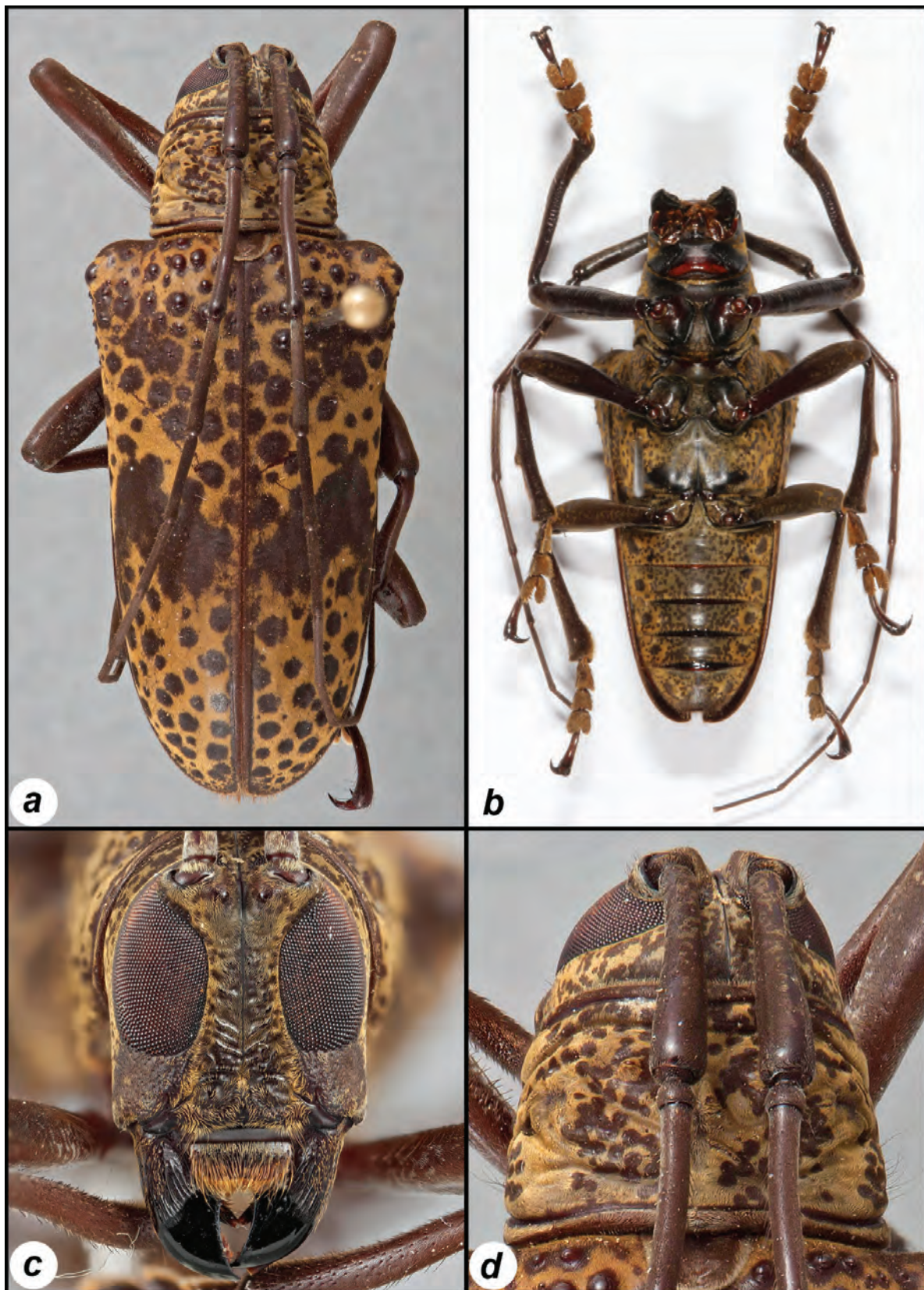


Figure 1. *Jamesia ramirezi*, sp. nov. **a)** Holotype female, dorsal habitus. **b)** Allotype male, ventral habitus. **c)** Holotype female, close-up of head. **d)** Holotype female, close-up of pronotum.

Male. Length 20.0–27.0 mm (measured from vertex to elytral apices), width 8.5–11.1 mm (measured across humeri). Similar to female except antennae 1.5 times longer than body; antennal tubercles armed at apex with short, blunt horn; pronotum widest behind middle, about 1.3 times as wide as long; elytra about 4 times as long as pronotal length; larger specimen with profemora transversely rugose; fifth sternite without a median triangular impression.

Type Material. Holotype, female (Fig. 1a, c-d), “Estación Pitilla, 9 km S. Sta. Cecilia, Prov. Guanacaste, Costa Rica. 700m, Abr. 1994. C. Moraga, LN 330200_380200 #2841” (INBC). Allotype, male (Fig. 1b), “Costa Rica, Limón, Liverpool, Reserva Veragua Rainforest. 430m.s.n.m. 9°55'35, 7" N - 83°11'27,9"W, 10/VI/2010, Rolando Ramírez Campos leg., #4503” (MNCR). Six paratypes: one male, same data as allotype except “18/VII/2011, #4757” (RRCC); one male, same data as allotype except “22/I/2009, #4185” (RRCC); one female, same data as holotype except “Mar 1994. #2804” (INBC); one male, same data as holotype except “P.N. Guanacaste, Ene 1994, #2563” (ENPC); one male, same data as holotype except “P.N. Guanacaste, May 1994. #2999” (INBC); one female, “Est. Hitoy Cerere, 100m. R. Cerere, Res. Biol. Hitoy Cerere, Prov. Limón, Costa Rica, 27 jun a 22 jul 1992, K. Taylor, L-N 184200, 643300” (INBC).

Etymology. *Jamesia ramirezi*, sp. nov. is named for Rolando Ramírez Campos, for his collaboration and who collected part of the type series. The epithet is a noun in the genitive case.

Diagnosis and Remarks. This species is distinguished from its congeners by the combination of the following characters: antennomeres I, II, basal 5/6 of III, and basal 2/3 of IV with grayish pubescence, distinctly lighter than V-XI, which are uniformly dark brown; and elytra with red-orange pubescence, with small and moderately sized, dark brown maculae outlined with ring of white pubescence. Swift et al. (2010) listed three species of *Jamesia* from Costa Rica. *Jamesia ramirezi*, sp. nov. is described from eight specimens: five males and three females. Nothing is known about the habitat and behavior of this species; however, all known specimens were collected in Costa Rica, above 100 m elevation (three specimens at 700 m elevation).

Peritrox Bates, 1865: 313 (Lamiinae: Onciderini)

Type species. *Peritrox denticollis* Bates, 1865 (monotypy).

The genus *Peritrox* currently contains four described species. Nearns et al. (2011) provided color photographs of three species of this genus, including two type specimens. The following key was adapted from Dillon and Dillon (1945) and treats all currently known species of *Peritrox* including one new species described herein (the species *Peritrox insulatus* Rodrigues and Mermudes, 2011, is transferred to the genus *Ischiocentra* Thomson herein).

1. Eye with lower lobe less than twice the height of gena; elytra with irregular, vermicular, fulvous maculae outlined with thin whitish pubescence, interspaces glabrous (Brazil).....
.....*P. vermiculatus* Dillon and Dillon, 1945
- Eye with lower lobe at least twice the height of gena; elytra without vermicular fulvous maculae 2
- 2(1). Elytra with fulvous or red-range pubescent maculae..... 3
- Elytra with only dull reddish pubescent maculae (Argentina, Bolivia, Brazil, Paraguay, Uruguay).....*P. denticollis* Bates, 1865
- 3(2). Antennomeres more or less uniformly dark brown; elytra fulvous, with irregularly shaped dark brown or black maculae outlined with thin grayish pubescence..... 4
- Antennomeres I, II, basal 5/6 of III, and basal 2/3 of IV with grayish pubescence, distinctly lighter than V-XI, which are uniformly dark brown; elytra with red-orange pubescence, with small and moderately-sized, dark brown maculae outlined with ring of white pubescence (French Guiana)*P. marcelae*, sp. nov.

- 4(3). Front entirely ochraceous pubescent, more densely pubescent laterally; elytra with fulvous pubescence predominating, a broad, unbroken, hoary fascia placed partly anterior to middle of its length (Brazil, Ecuador, French Guiana, Peru).... *P. perbra* Dillon and Dillon, 1945
- Front variegated with white and fulvous pubescence, fulvous vittate laterally; elytra with gray pubescence predominating, hoary fascia broken, placed behind middle of length (Bolivia, Brazil, Paraguay)..... *P. nigromaculata* Aurivillius, 1920

***Peritrox marcelae* Nearns and Tavakilian, sp. nov.**

(Figures 2a–d)

Description. Male. Length 16.0–17.0 mm (measured from vertex to elytral apices), width 5.5–6.0 mm (measured across humeri). Habitus as in Fig. 2a. General form elongate-oblong, moderate-sized. Integument dark brown, antennomeres I, II, basal 5/6 of III, and basal 2/3 of IV with grayish pubescence, distinctly lighter than V–XI, which are uniformly dark brown; elytra with red-orange pubescence, with dense field of small and moderately-sized, dark brown maculae outlined with ring of white pubescence; legs with grayish pubescence; legs with grayish pubescence.

Head with frons distinctly elongate, a little wider than width of one lower eye lobe (as in Fig. 2c). Eyes with lower lobes large, oblong; narrowest area connecting upper and lower eye lobes about 6 ommatidia wide. Genae roughly subquadrate, about half as tall as lower eye lobes.

Antennae about 2 times longer than body; antennal tubercles prominent, narrowly separated, contiguous at base; tubercles armed at apex with short blunt horn; scape robust, gradually expanded to apex; antennomeres III, X, and XI slightly curved. Antennal formula based on antennomere III: scape=0.55; II=0.08; III=1; IV=0.72; V=0.6; VI=0.54; VII=0.5; VIII=0.48; IX=0.47; X=0.51; XI=0.77.

Pronotum subcylindrical, slightly wider at base, transverse, about 1.5 times as wide as long, sides irregular, with a small, blunt protuberance each side behind middle (Fig. 2a, d); disk with five feebly elevated tubercles, median tubercle oval, lateral tubercles slightly longer, elongate; disk with 8–10 fine punctures at basal transverse sulcus.

Scutellum transverse, apex rounded.

Elytra about 2 times as long as width at humeri (Fig. 2a), nearly 4 times as long as pronotal length, about 1.3 times broader basally than pronotum at widest (at base); lateral margins nearly straight, sides roughly parallel, gradually rounded to apices at apical 1/3, apices jointly rounded; basal 1/3 of elytra with moderate punctation, surface finely punctate, with mix of shallow and deep punctures; humeri prominent, anterior margin arcuate, angle with moderate sized, obtuse tubercle.

Venter with procoxae large, globose, not uncate; narrowest area of prosternal process between procoxae about 1/6 as wide as procoxal cavity; apex of prosternal process subtriangular. Mesosternal process about as wide as mesocoxal cavity; mesosternal process subtruncate-rounded. Fifth sternite slightly longer than IV.

Legs moderate in length; femora robust; metafemora clavate apically; tibiae slightly expanded apically; metafemora about 1/3 as long as elytra.

Female. Length 19.5–24.0 mm (measured from vertex to elytral apices), width 7.5–9.0 mm (measured across humeri). Similar to male except antennae about 1.5 times as long as body; elytra about 1.5 times broader basally than pronotum at widest (at middle).

Type Material. Holotype, male (Fig. 2a, c), “Montagne de Kaw, 11.XII.82 pk 35 P.L., P. Debost leg.” (MNHN). Allotype, female (Fig. 2b), “05/I/2011 lumière Piste Kapiri RN2 PK125, J-L Giuglaris leg.” (PHDC). Four paratypes: one male (Fig. 2d), “Guyane, 05/I/2008 lumière piste de Bélizon PK15+17, J-L Giuglaris leg.” (JLGC); one male, “Guyane, 06/IX/ 2011 lumière ZA Wayabo Matiti, J-L Giuglaris leg.” (JLGC); one female, “Guyane, 10/I/1994 lumière route de Kaw PK38, J-L Giuglaris leg.” (JLGC); one female, “Guyane, 01/IV/2003 lumière piste de Bélizon PK15+12” (JLGC).

Etymology. We are pleased to name this species in honor of Marcela Laura Monné, for her friendship and many contributions to the study of Neotropical Cerambycidae. The epithet is a noun in the genitive case.

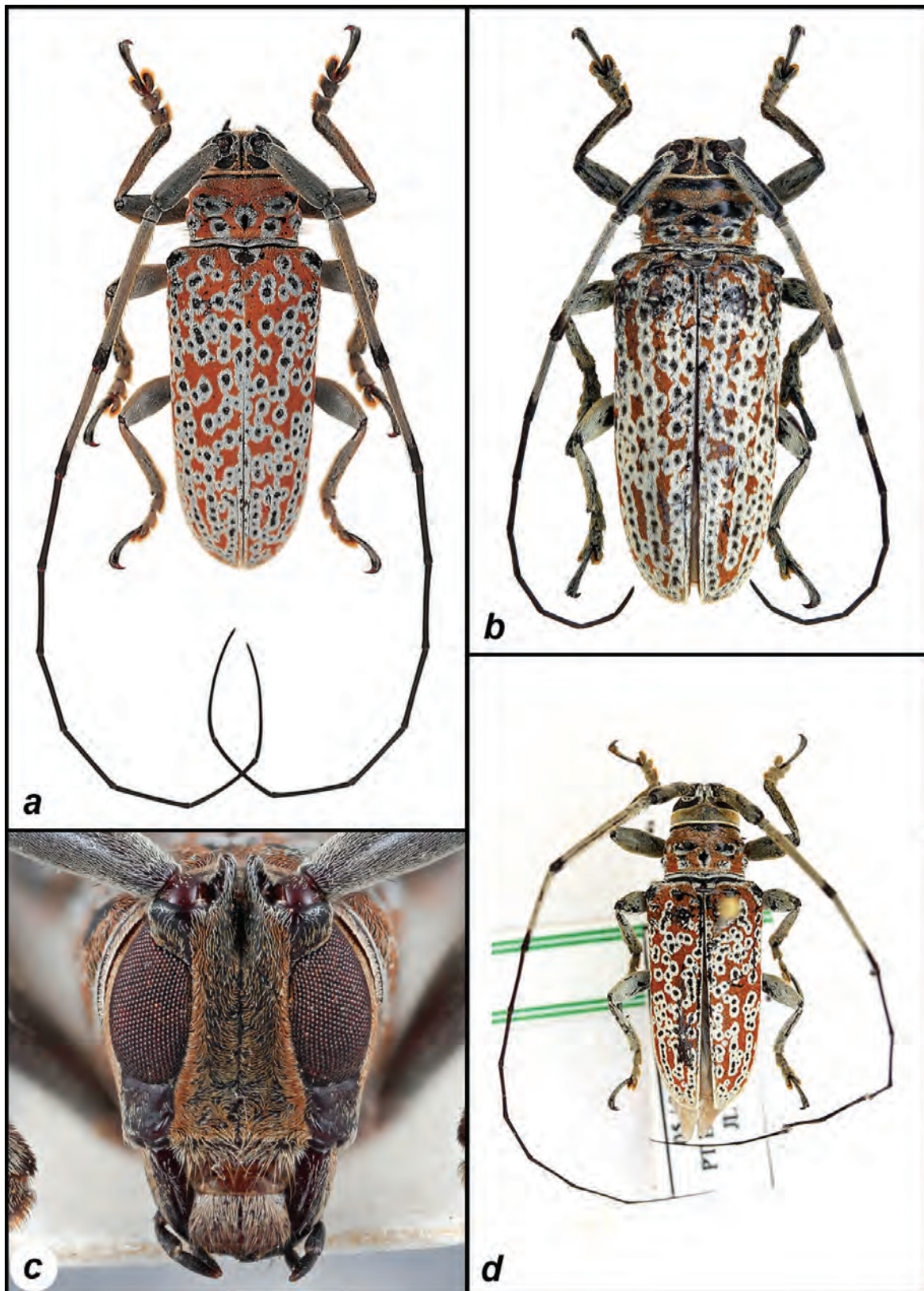


Figure 2. *Peritrox marcelae*, sp. nov. a) Holotype male, dorsal habitus. b) Allotype female, dorsal habitus. c) Holotype male, close-up of head. d) Paratype male, dorsal habitus.

Diagnosis and Remarks. This species is distinguished from its congeners by the combination of the following characters: antennomeres I, II, basal 5/6 of III, and basal 2/3 of IV with grayish pubescence, distinctly lighter than V–XI, which are uniformly dark brown; and elytra with red-orange pubescence, with small and moderately sized, dark brown maculae outlined with ring of white pubescence. *Peritrox marcelae*, sp. nov. is described from six specimens: three males and three females. Nothing is known about the habitat and behavior of this species; however, all known specimens were collected in French Guiana, at light.

***Touroultia* Nearns and Tavakilian, gen. nov. (Lamiinae: Onciderini)**

(Figures 4a–d, 5a–d)

Type species. *Touroultia lordi*, sp. nov., here designated.

Description. General form elongate-ovate, robust, small to moderate-sized. Head with frons elongate, about 1.5 times width of lower eye lobe. Eyes with lower lobes large, oblong. Genae distinctly shorter than lower eye lobes. Antennae distinctly longer than body; antennal tubercles prominent, moderately separated; scape robust, clavate; antennomere III longest. Pronotum subcylindrical, transverse, sides feebly sinuate; disk with three tubercles; disk with one basal transverse sulcus, and a more distinct oblique sulcus laterally which continues down the side. Scutellum transverse, apex rounded. Elytra with lateral margins slightly sinuate, gradually rounded to apices at apical 1/3; humeri prominent. Legs moderate in length; profemora robust; meso- and metafemora clavate apically; tibiae slightly expanded apically; metafemora about 1/3 as long as elytra.

The following key treats all currently known species including two new species described herein.

1. Genae roughly transverse, about 1/3 as tall as lower eye lobes; elytra with basal gibbositities moderately elevated..... **2**
- Genae roughly subquadrate, about 1/2 as tall as lower eye lobes; elytra with basal gibbositities feebly elevated (Brazil) ***T. obscurella* (Bates, 1865), new combination**
- 2(1). Base of antennomeres IV–XI with grayish or pale testaceous pubescence; base of elytra with surface coarsely, moderately to deeply, granulate-punctate (French Guiana) ***T. lordi*, sp. nov.**
- Base of antennomeres IV–XI with golden-yellow pubescence; base of elytra with surface finely, shallowly, granulate-punctate (Ecuador) ***T. swifti*, sp. nov.**

Etymology. *Touroultia*, gen. nov. is named for Julien Touroult, with appreciation of his friendship and collaboration. The gender is feminine.

Diagnosis and Remarks. This genus closely resembles *Priscatoides* Dillon and Dillon, 1945 (Fig. 3a–d) but can be distinguished by the combination of the following characters: genae distinctly shorter than lower lobes, from 1/3 to 1/2 as tall (genae slightly shorter than lower eye lobes in *Priscatoides*); frons distinctly elongate and narrow, about as wide as width of 1 to 1.5 lower eye lobes (frons about as wide as width of two lower eye lobes in *Priscatoides*); pronotum subcylindrical in *Touroultia* (slightly narrower at apex in *Priscatoides*); elytra about 3.75 times longer than pronotal length (about 4.3 times longer in *Priscatoides*).

***Touroultia lordi* Nearns and Tavakilian, sp. nov.**

(Figures 4a, c)

Description. Male. Length 14.0–14.5 mm (measured from vertex to elytral apices), width 6.0–6.25 mm (measured across humeri). Habitus as in Fig. 4a. General form elongate-ovate, moderate-sized.

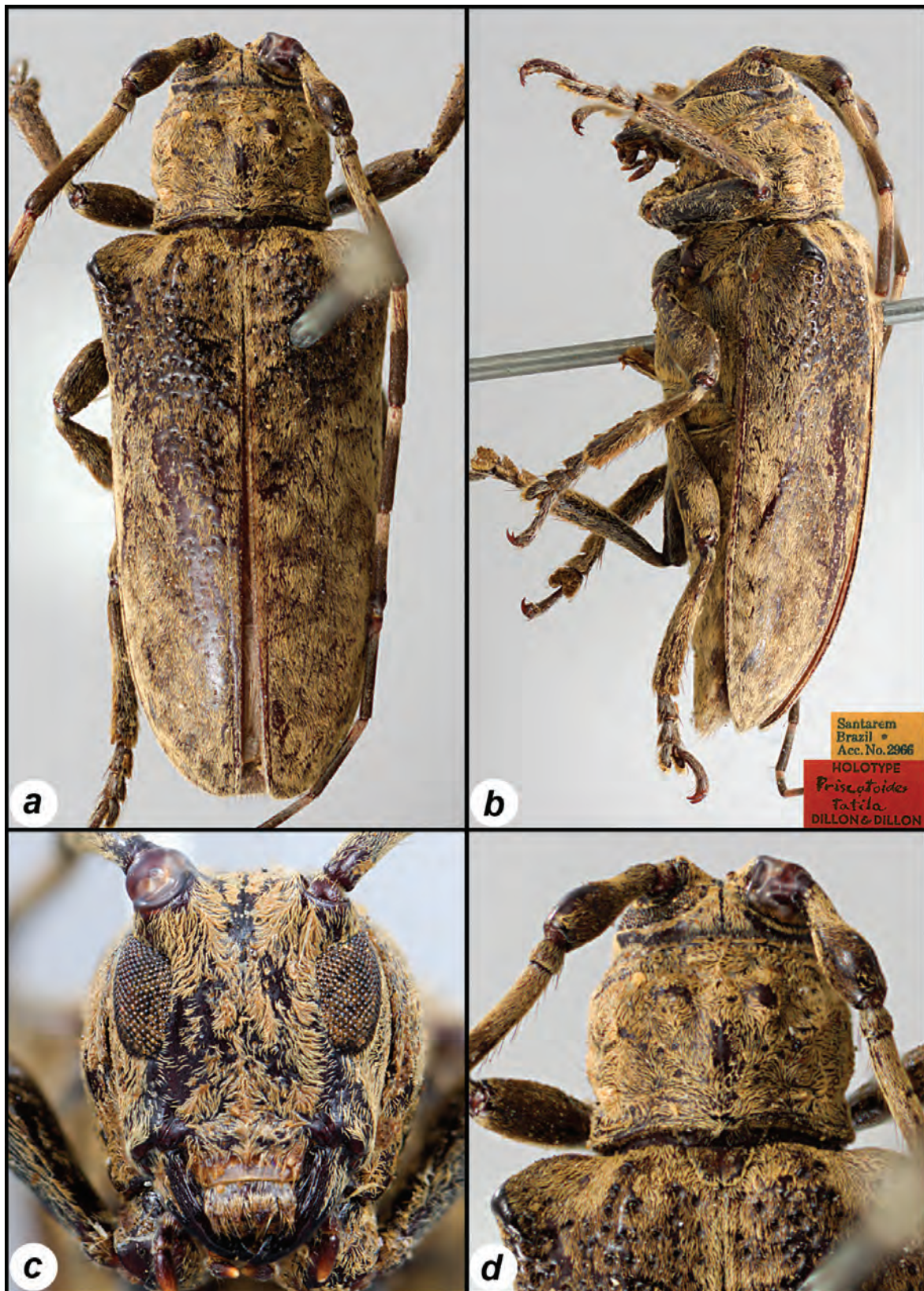


Figure 3. *Priscatoides tatila* Dillon and Dillon, 1945, holotype female. **a)** Dorsal habitus. **b)** Lateral habitus with original labels. **c)** Close-up of head. **d)** Close-up of pronotum.

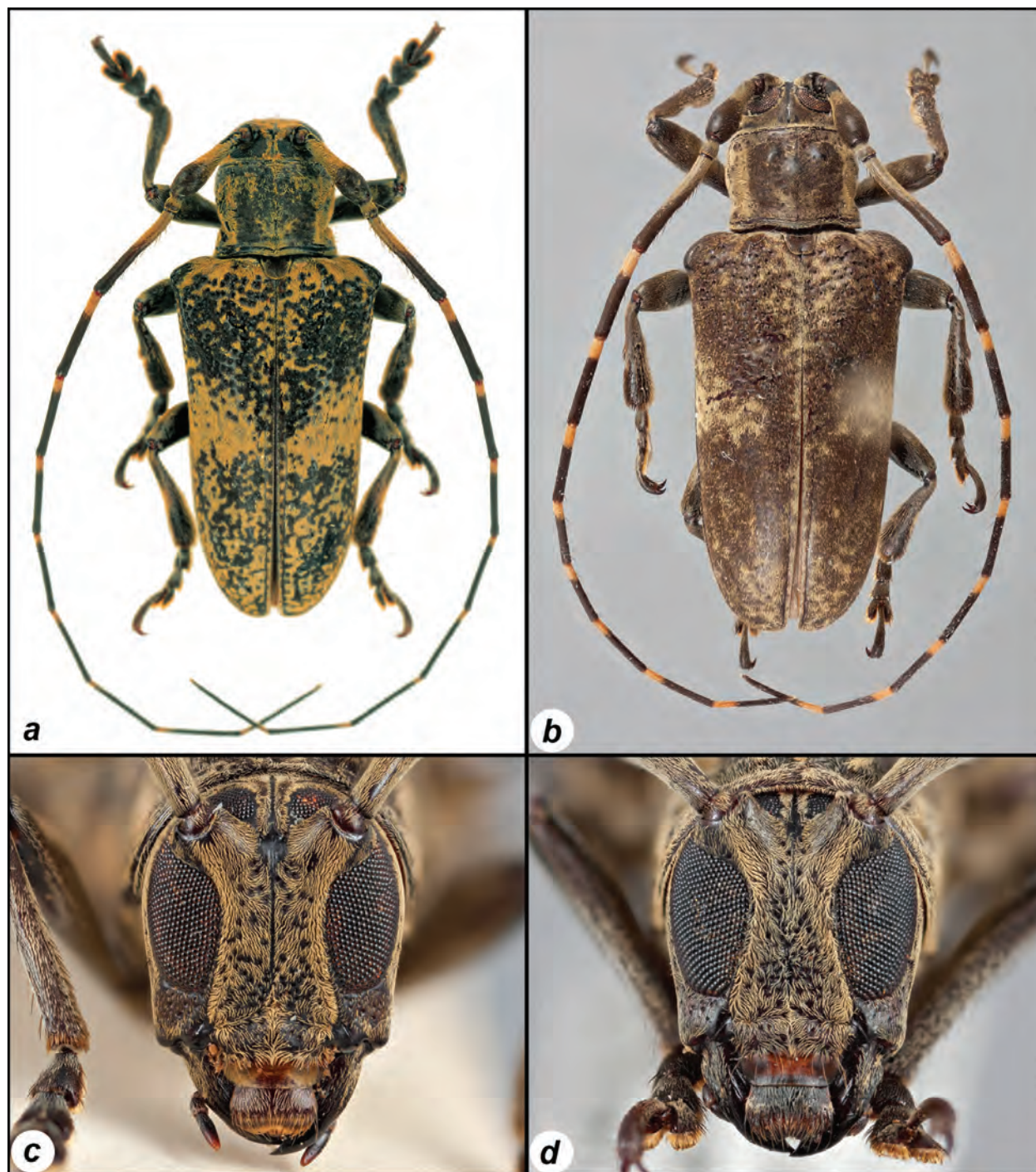


Figure 4. Two species of *Touroultia*, gen. nov. **a)** *T. lordi*, sp. nov., holotype male, dorsal habitus. **b)** *T. swifti*, sp. nov., holotype male, dorsal habitus. **c)** *T. lordi*, sp. nov., holotype male, close-up of head. **d)** *T. swifti*, sp. nov., holotype male, close-up of head.

Integument dark brown or black, with pale testaceous and dark brown variegated pubescence; base of antennomeres with grayish or pale testaceous pubescence; elytra dark brown pubescence, with two irregularly-shaped, distinct regions of pale testaceous pubescence near middle.

Head with frons elongate, about 1.5 times width of lower eye lobe (as in Fig. 4c). Eyes with lower lobes large, oblong; narrowest area connecting upper and lower eye lobes about 4 ommatidia wide.

Genae roughly subquadrate, about 1/3 as tall as lower eye lobes.

Antennae distinctly longer than body; antennal tubercles prominent, moderately separated; tubercles unarmed at apex; scape robust, clavate; antennomere III sinuate. Antennal formula based on antennomere III: scape=0.84; II=0.15; III=1; IV=0.85; V=0.76; VI=0.73; VII=0.74; VIII=0.74; IX=0.77; X=0.79; XI=0.87.

Pronotum subcylindrical, transverse, about 1.25 times as wide as long, sides feebly sinuate; disk with three moderately elevated tubercles, median tubercle oval, lateral tubercles more prominent; one basal transverse sulcus, and a more distinct oblique sulcus laterally which continues down the side.

Scutellum transverse, apex rounded.

Elytra about 1.6 times as long as width at humeri (Fig. 4a), about 3.75 times as long as pronotal length, about 1.6 times broader basally than pronotum at widest (at base); lateral margins slightly sinuate, gradually rounded to apices at apical 1/3, apices individually rounded; base of each elytron with a moderate gibbosity; basal 1/3 of elytra with surface coarsely, moderately to deeply, granulate-punctate; humeri prominent, anterior margin oblique, angle with moderate sized, obtuse tubercle.

Venter with procoxae large, globose, with small, obtuse tubercle; narrowest area of prosternal process between procoxae, about 1/5 as wide as procoxal cavity; apex of prosternal process subtriangular. Mesosternal process about 0.75 times as wide as mesocoxal cavity; mesosternal process feebly emarginate. Fifth sternite slightly longer than IV, apex feebly emarginate.

Legs moderate in length; profemora robust; meso- and metafemora clavate apically; tibiae slightly expanded apically; metafemora about 1/3 as long as elytra.

Female. Length 17.0 mm (measured from vertex to elytral apices), width 7.5 mm (measured across humeri). Similar to male except antennae about 1.3 times as long as body; procoxae without tubercle; elytra about 1.75 times broader basally than pronotum at widest (at middle); fifth sternite about 1.5 times as long as IV, with a median triangular impression.

Type Material. Holotype, male (Fig. 4a, c), "Piste Coralie pk 12, 8 Janvier 1988, Guyane, Piégeage lumineux, Odette Baloup leg." (MNHN). Allotype, female, "Guyane F. Route de Kaw, PK 33, battage, 19-27-XII-2003, Touroult leg." (JTTC). Two paratypes: one male, "French Guiana, PK 37-40, Rte de Kaw, Jan 25-Feb 1, 1995, E. Giesbert, F. Hovore" (ENPC); one male, "Montagne des Chevaux GF, piège à vitre, 08/II/2009, P.-H. Dalens leg." (PHDC).

Etymology. We take pleasure in naming this species for Nathan Patrick Lord, for his friendship and companionship on many collecting trips. The epithet is a noun in the genitive case.

Diagnosis and Remarks. This species is distinguished from its congeners by the combination of the following characters: base of antennomeres IV-XI with grayish or pale testaceous pubescence; base of elytra with surface coarsely, deeply, granulate-punctate genae roughly transverse, about 1/3 as tall as lower eye lobes; elytra with basal gibbositities strongly elevated. *Touroultia lordi*, sp. nov. is described from four specimens: three males and one female. Nothing is known about the habitat and behavior of this species; however, the holotype specimen was collected at light and the allotype specimen was collected beating vegetation.

***Touroultia obscurella* (Bates, 1865), new combination**

(Figures 5a–d)

Hypselomus obscurellus Bates, 1865: 169.

Type locality: Brazil, Pará: Óbidos. (MNHN). Distribution: Brazil (Pará).

Hypsioma obscurella (Bates, 1865); Lacordaire 1872: 676; Dillon and Dillon 1946: 206; Martins and Galileo 1990: 56.

Hypsioma ? *obscurella*; Breuning 1961: 208 (cat.).

Description. Male? Length 11.0 mm (measured from vertex to elytral apices), width 4.9 mm (measured across humeri). Habitus as in Fig. 5a. General form elongate-ovate, small-sized. Integument ferrugine-

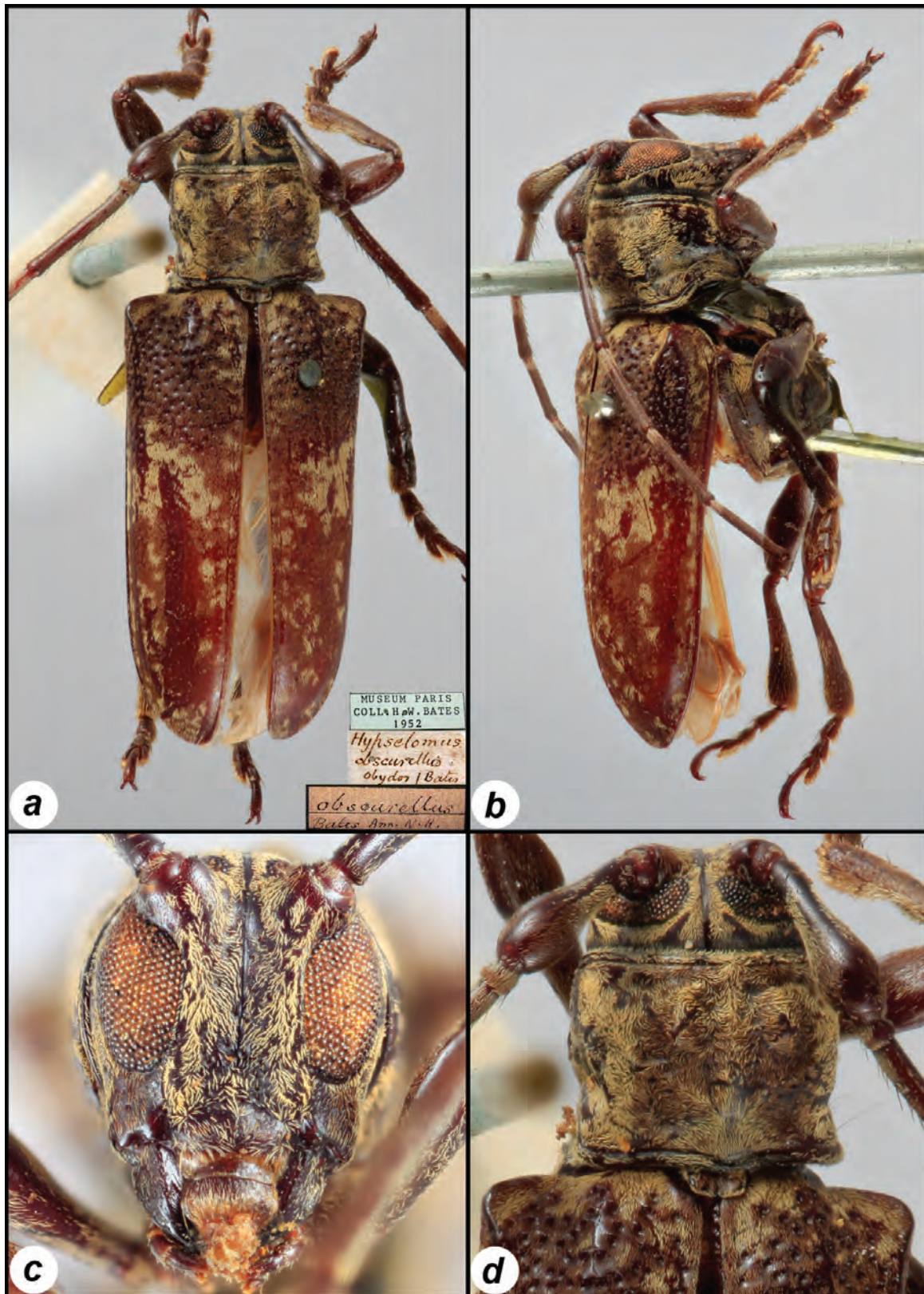


Figure 5. *Touroultia obscurella* (Bates, 1865), holotype male? a) Dorsal habitus with original labels. b) Lateral habitus. c) Close-up of head. d) Close-up of pronotum.

ous or dark brown, with grayish and pale testaceous pubescence; base of antennomeres with grayish pubescence; elytra with predominantly dark brown pubescence, with two feebly defined regions of pale testaceous pubescence near middle.

Head with frons elongate, about 1.5 times width of lower eye lobe (as in Fig. 5d). Eyes with lower lobes large, oblong; narrowest area connecting upper and lower eye lobes about 4 ommatidia wide. Genae roughly subquadrate, about 1/2 as tall as lower eye lobes.

Antennae about 1.3 times longer than body (estimated); tubercles prominent, moderately separated; tubercles unarmed at apex; scape robust, clavate; antennomere III sinuate. Antennal formula based on antennomere III: scape=0.73; II=0.13; III=1; IV=0.82; V=0.7; VI=0.57 (specimen damaged, antennomeres VII-XI missing).

Pronotum subcylindrical, transverse, about 1.25 times as wide as long, sides feebly sinuate; disk with three moderately elevated tubercles, median tubercle oval, lateral tubercles more prominent; one basal transverse sulcus, and a more distinct oblique sulcus laterally which continues down the side.

Scutellum transverse, apex rounded.

Elytra about 1.85 times as long as width at humeri (Fig. 5b), about 3.75 times as long as pronotal length, about 1.6 times broader basally than pronotum at widest (at base); lateral margins slightly sinuate, gradually rounded to apices at apical 1/3, apices individually rounded; base of each elytron with a moderate gibbosity; basal 1/3 of elytra with dense punctation, surface finely granulate-punctate; humeri prominent, anterior margin oblique, angle with moderate sized, obtuse tubercle.

Venter with procoxae large, globose, not uncate. Abdomen unavailable for study (specimen damaged).

Legs moderate in length; profemora robust; meso- and metafemora clavate apically; tibiae slightly expanded apically; metafemora about 1/3 as long as elytra.

Female. Unknown.

Material Examined. Holotype, male? (Fig. 5a-d), "*Hypselomus obscurellus*, Obydos / Bates; obscurellus Bates Am. N.H.; Museum Paris Coll. H.W. Bates 1952" (MNHN).

Diagnosis and Remarks. This species is distinguished from its congeners by the combination of the following characters: genae roughly subquadrate, about 1/2 as tall as lower eye lobes; elytra with basal gibbositities feebly elevated. *Touroultia obscurella* (Bates, 1865) is described from a single specimen collected at "Obydos" (Brazil, Pará: Óbidos) and originally described in the genus *Hypselomus* Perty, 1832 (Onciderini). Although the original description indicates the holotype specimen is male, this remains unconfirmed due to specimen damage. Breuning (1961) indicated doubt about the placement of this species in the genus *Hypsioma* Audinet-Serville, 1835 (Onciderini).

***Touroultia swifti* Nearns and Tavakilian, sp. nov.**

(Figures 4b, d)

Description. Male. Length 14.0 mm (measured from vertex to elytral apices), width 5.8 mm (measured across humeri). Habitus as in Fig. 4b. General form elongate-ovate, moderate-sized. Integument dark brown or black, with pale testaceous and dark brown variegated pubescence; pedicel, base of scape and antennomere III with grayish pubescence; base of antennomeres IV-XI with golden-yellow pubescence; elytra with predominantly dark brown pubescence, with two feebly defined regions of pale testaceous pubescence near middle.

Head with frons elongate, about width of 1 lower eye lobe (as in Fig. 4d). Eyes with lower lobes distinctly large, oblong; narrowest area connecting upper and lower eye lobes about 4 ommatidia wide. Genae transverse, about 1/3 as tall as lower eye lobes.

Antennae about 1.5 times longer than body; antennal tubercles prominent, moderately separated; tubercles armed at apex with short blunt tooth; scape robust, clavate; antennomere III sinuate. Antennal formula based on antennomere III: scape=0.67; II=0.16; III=1; IV=0.85; V=0.82; VI=0.7; VII=0.55; VIII=0.56; IX=0.63; X=0.65; XI=0.71.

Pronotum subcylindrical, transverse, about 1.25 times as wide as long, sides feebly sinuate; disk with three moderately elevated tubercles, median tubercle oval, lateral tubercles more prominent; one basal transverse sulcus, and a more distinct oblique sulcus laterally which continues down the side.

Scutellum transverse, apex rounded.

Elytra about 1.6 times as long as width at humeri (Fig. 4b), about 3.75 times as long as pronotal length, about 1.6 times broader basally than pronotum at widest (at base); lateral margins slightly sinuate, gradually rounded to apices at apical 1/3, apices individually rounded; base of each elytron with a moderately-elevated gibbosity; basal 1/3 of elytra with dense punctation, surface finely granulate-punctate; humeri prominent, anterior margin oblique, angle with moderate sized, obtuse tubercle.

Venter with procoxae large, globose, with small, acute tubercle; narrowest area of prosternal process between procoxae distinctly narrow, about 1/10 as wide as procoxal cavity; apex of prosternal process subtriangular. Mesosternal process about 0.75 times as wide as mesocoxal cavity; mesosternal process feebly emarginate. Fifth sternite slightly longer than IV, apex feebly emarginate.

Legs moderate in length; profemora robust; meso- and metafemora feebly clavate apically; tibiae slightly expanded apically; metafemora about 1/3 as long as elytra.

Female. Length 17.0 mm (measured from vertex to elytral apices), width 7.5 mm (measured across humeri). Similar to male except antennae about 1.25 times as long as body; procoxae without tubercle; fifth sternite about 1.5 times as long as IV, with a median triangular impression.

Type Material. Holotype, male (Fig. 4b), "Ecuador, Napo Prov., 24km E. Atahualpa, 450m, Sept 20-22, 1996, E. Giesbert, coll." (EFGC). Allotype, female, "Ecuador, Napo Pr., 1 km W Coca, 08 Oct 1997, F.T. Hovore, coll." (ENPC).

Etymology. We are pleased to name this species for Ian Patrick Swift, with appreciation of his friendship, encouragement, and camaraderie in the field. The epithet is a noun in the genitive case.

Diagnosis and Remarks. This species is distinguished from its congeners by the combination of the following characters: genae transverse, about 1/3 as tall as lower eye lobes; base of antennomeres IV-XI with golden-yellow pubescence; base of elytra with surface finely, shallowly, granulate-punctate; elytra with basal gibbositities moderately elevated. *Touroultia swifti*, sp. nov. is described from two specimens: one male and one female. Nothing is known about the habitat and behavior of this species; however, both specimens were collected in Ecuador and the male specimen was collected at 450 m elevation.

***Trestoncideres* Martins and Galileo, 1990: 87 (Lamiinae: Onciderini)**

Type species. *Trestoncideres laterialba* Martins and Galileo, 1990 (monotypy and original designation).

The genus *Trestoncideres* currently contains two described species. Nearns et al. (2011) provided color photographs of both species, including the holotype specimen of *Trestoncideres laterialba* Martins and Galileo, 1990 and a paratype specimen of *Trestoncideres albiventris* Martins and Galileo, 2005. The following key treats all currently known species of *Trestoncideres* including one new species described herein.

1. Antennae, pronotum, and elytra with whitish and gold pubescence; pronotum with a small, acute protuberance each side behind middle; scutellum with central 1/2 glabrous, outer margins fringed with grayish or whitish pubescence; apical 1/3 of elytra slightly darker (Brazil, French Guiana)..... ***T. santossilvai*, sp. nov.**
- Antennae, pronotum, and elytra without grayish-white and gold pubescence; pronotum without acute protuberances each side; scutellum pubescent, with at most a small glabrous area at center; apical 1/3 of elytra not slightly darker **2**

- 2(1). Base of each elytron with moderately raised gibbosity; elytral gibbositities with orange pubescence, together forming an elongate-cordate region; elytra (excluding gibbositities) with nearly uniform ochraceous or light brown pubescence (Bolivia).....*T. albiventris* Martins and Galileo, 2005
- Elytra with nearly uniformly pale orange pubescence (Brazil, Costa Rica, French Guiana, Suriname).....*T. laterialba* Martins and Galileo, 1990

***Trestoncideres santossilvai* Nearn and Tavakilian, sp. nov.**

(Figures 6a–d)

Description. Male. Length 12.0–13.0 mm (measured from vertex to elytral apices), width 3.5–4.0 mm (measured across humeri). Habitus as in Fig. 6a. General form elongate-oblong, small to moderate-sized. Integument generally dark brown or black, with whitish, gold, and ochraceous pubescence; mesepisternum, mesepimeron, metepisternum with dark brown and gold pubescence; metasternum with distinct vitta formed by dense, white pubescence, about as wide as metepisternum, not reaching margin with metepisternum (Fig. 6b); lateral margin of metasternum with dark brown and gold pubescence; remaining portions of venter and femora with whitish pubescence; apical 1/3 of elytra slightly darker than basal 2/3.

Head with frons roughly subquadrate, slightly transverse, about 3 times width of lower eye lobe (as in Fig. 6c). Eyes with lower lobes moderate-sized, ovate; narrowest area connecting upper and lower eye lobes about 2–3 ommatidia wide. Genae elongate, about as tall as lower eye lobes. Antennae about 1.25 times longer than body; antennal tubercles prominent, widely separated; tubercles armed at apex with short blunt horn; scape robust, clavate; antennomere III curved, slightly sinuate. Antennal formula based on antennomere III: scape=0.85; II=0.29; III=1; IV=0.76; V=0.72; VI=0.65; VII=0.63; VIII=0.5; IX=0.53; X=0.47; XI=0.48.

Pronotum roughly subcylindrical, slightly narrower at base, transverse, about 1.4 times as wide as long, sides irregular, with a small, acute protuberance each side behind middle (Fig. 6d); disk with median, oval, glabrous region at basal half.

Scutellum transverse, apex rounded; outer margins fringed with whitish pubescence, central 1/2 glabrous.

Elytra about 2 times as long as width at humeri (Fig. 6a), about 3.3 times as long as pronotal length, about 1.25 times broader basally than pronotum at widest (at base); lateral margins nearly straight, gradually rounded to apices at apical 1/3, apices jointly rounded; base of each elytron with a feeble gibbosity; basal 1/3 of elytra with moderate to dense punctation, surface granulate-punctate; humeri slightly prominent, anterior margin arcuate, angle with small, obtuse tubercle.

Venter with procoxae large, globose, not uncate; narrowest area of prosternal process between procoxae about 1/4 as wide as procoxal cavity; apex of prosternal process subtriangular. Mesosternal process about 1/3 as wide as mesocoxal cavity; mesosternal process subtruncate. Fifth sternite about 1.5 times as long as IV, apex feebly emarginate.

Legs short in length; pro- and mesofemora robust; metafemora clavate apically; tibiae expanded apically; metafemora about 1/4 as long as elytra.

Female. Length 12.0–14.0 mm (measured from vertex to elytral apices), width 4.0–4.5 mm (measured across humeri). Similar to male except antennae shorter, surpassing elytral apex at antennomere X; fifth sternite about 3 times as long as IV, with a median triangular impression.

Type Material. Holotype, male (Fig. 6a, c–d), “Piste de Belizon, pk 24, 19 septembre 1992, Guyane, piégeage lumineux, Michel Duranton leg.” (MNHN). Allotype, female (Fig. 6b), “Brazil: Amazonas, Rio Taruma Mirim, 20 km nw Manaus, 02Mar1979, 02°53’S 060°07’W; Black water inundation forest canopy fogged with Pyrethrum sample #44; Montgomery, Irwin, Schimmel, Krischik, Date, Bacon colls.”

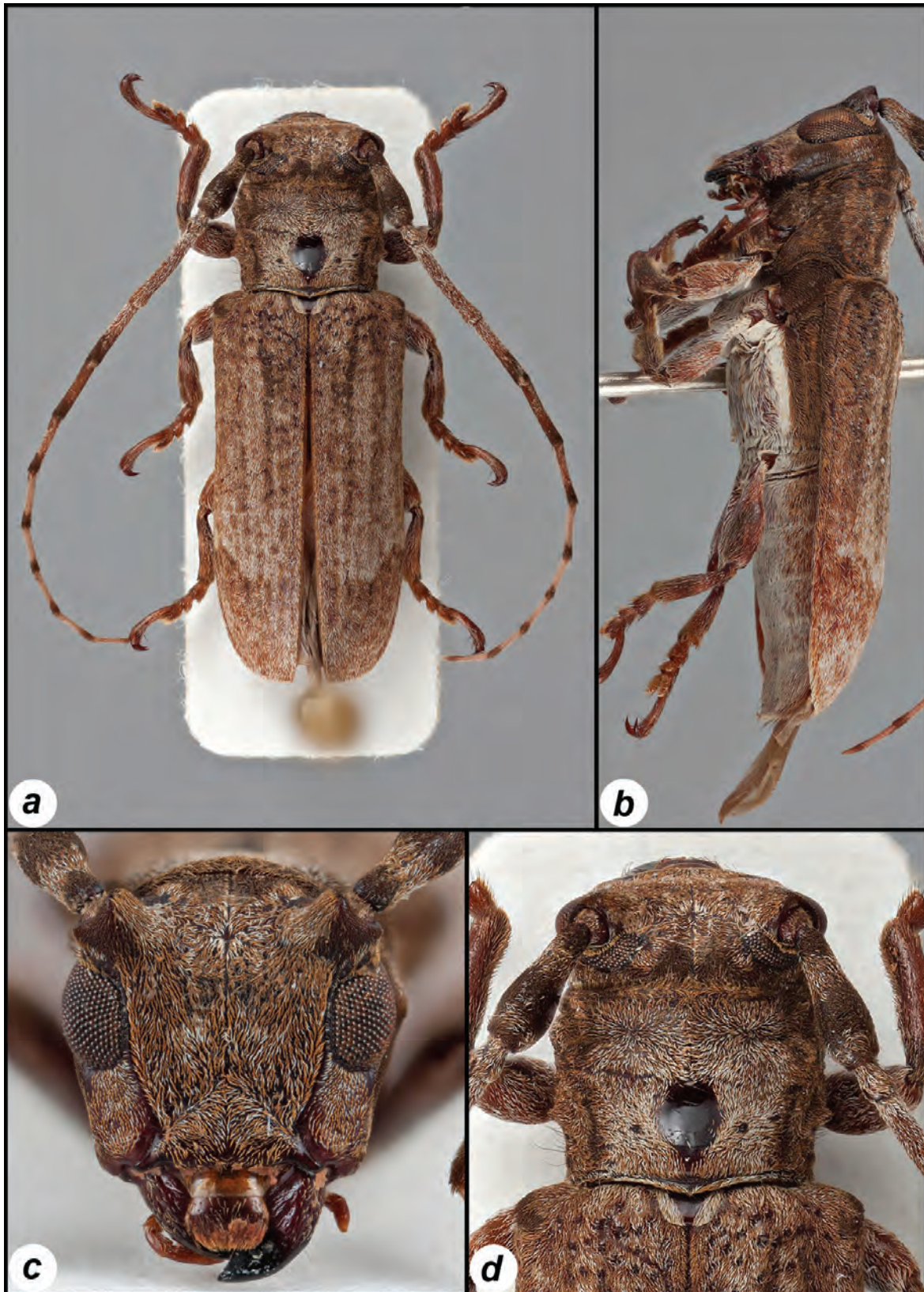


Figure 6. *Trestoncideres santossilvai*, sp. nov. **a)** Holotype male, dorsal habitus. **b)** Allotype female, lateral habitus. **c)** Holotype male, close-up of head. **d)** Holotype male, close-up of pronotum.

(USNM). Nine paratypes: one male, "Montsinery FRG Guyane, 24 Juillet 1985, Piégeage lumineux, P. Souka leg., 0489" (MNHN); one male, "Risque-tout (FRG pk19), 7 Mai 1983, Piégeage lumineux, P. Sarry leg., 0489" (MNHN); one female, "Route de Kaw pk 38, 13 Janvier 1986, Guyane, Piégeage lumineux, P. Gerdelat leg., 0489" (ENPC); one female, "Piste Risquetout pk 16, 28 Juin 1987, Guyane, Piégeage lumineux, Marc Thouvenot leg." (MNHN); one female, "Kaw PK 37, 20-VII-2001, PL" (DFPC); one male, "Guyane, 12/V/1991 lumière route de Kaw PK47, J-L Giuglaris leg." (PHDC); one male, "Guyane, 06/III/1997 lumière RD06 PK19 J-L Giuglaris leg." (JLGC); one male, "Guyane, 21/VII/1990 lumière Cacao J-L Giuglaris leg." (JLGC); one female, "Guyane, 15/II/2006 lumière RD06 PK38 J-L Giuglaris leg." (JLGC).

Etymology. We take pleasure in naming this species in honor of Antonio Santos-Silva, for his friendship, collaboration, and many contributions to the study of Neotropical Cerambycidae and Disteniidae. The epithet is a noun in the genitive case.

Diagnosis and Remarks. This species is distinguished from its congeners by the combination of the following characters: antennae, pronotum, and elytra with whitish and gold pubescence; pronotum with a small, acute protuberance each side behind middle; scutellum glabrous except for outer margins, which are fringed with whitish pubescence; and apical 1/3 of elytra slightly darker than basal 2/3. In addition, *T. santossilvai* can be distinguished from its congeners by the lateral margins of the metasterna not with dense white pubescence (in *T. albiventris* and *T. laterialba*, the dense white pubescence extends to the lateral margins of the metasterna, contiguous with metepisterna); outermost margin of metepisternum with dark brown and gold pubescence. *Trestoncideres santossilvai*, sp. nov. is described from 11 specimens: six males and five females. Nothing is known about the habitat and behavior of this species; however, all specimens from French Guiana were collected at light and the specimen from Brazil was collected by canopy fogging with pyrethrum.

Taxonomic Notes

Clavidesmus Dillon and Dillon, 1946: 293 (Lamiinae: Onciderini)

(Figures 7a–i)

Clavidesmus Dillon and Dillon, 1946: 293. Type species: *Eudesmus heterocerus* Buquet, 1852.
= *Orteguaza* Lane, 1958: 10 (Lamiinae: Apomecynini). Type species: *Orteguaza lichenigera* Lane, 1958, by original designation, **new synonymy**.

Discussion. Lane (1958) described the genus *Orteguaza* and included two species: *O. funeraria* (Fig. 7c) and *O. lichenigera* (Fig. 7f). Examination of the holotype specimens of *O. funeraria* and *O. lichenigera*, as well as the type specimens of all described species of *Clavidesmus* Dillon and Dillon [*C. chiccae* Giorgi, 1998 (Fig. 7a); *C. columbianus* Breuning, 1961 (Fig. 7b); *C. heterocerus* (Buquet, 1852) (Fig. 7d); *C. indistinctus* Dillon and Dillon, 1952 (Fig. 7e); *C. metallicus* (Thomson, 1868) (Fig. 7g); *C. monnei* Giorgi, 1998 (Fig. 7h); and *C. rubiginus* Dillon and Dillon, 1949 (Fig. 7i)] revealed are no characters to separate the two genera. Based on close morphological similarities, *Orteguaza* is here **synonymized** with *Clavidesmus*, creating two **new combinations**, *Clavidesmus funerarius* (Lane, 1958) and *Clavidesmus lichenigerus* (Lane, 1958). The known range of *Clavidesmus* is extended to Central America (Costa Rica, Honduras, Nicaragua, and Panama). We believe the genus *Clavidesmus* is over-split and in need of a taxonomic revision.

Ischiocentra Thomson, 1861: 382 (Lamiinae: Onciderini)

Ischiocentra Thomson, 1861: 382. Type species: *Ischiocentra clavata* Thomson, 1861.

Discussion. Rodrigues and Mermudes (2011) described *Peritrox insulatus* from a single female specimen collected on "Ilha Grande," Rio de Janeiro, Brazil. Nearn et al. (2011) provided color photographs of

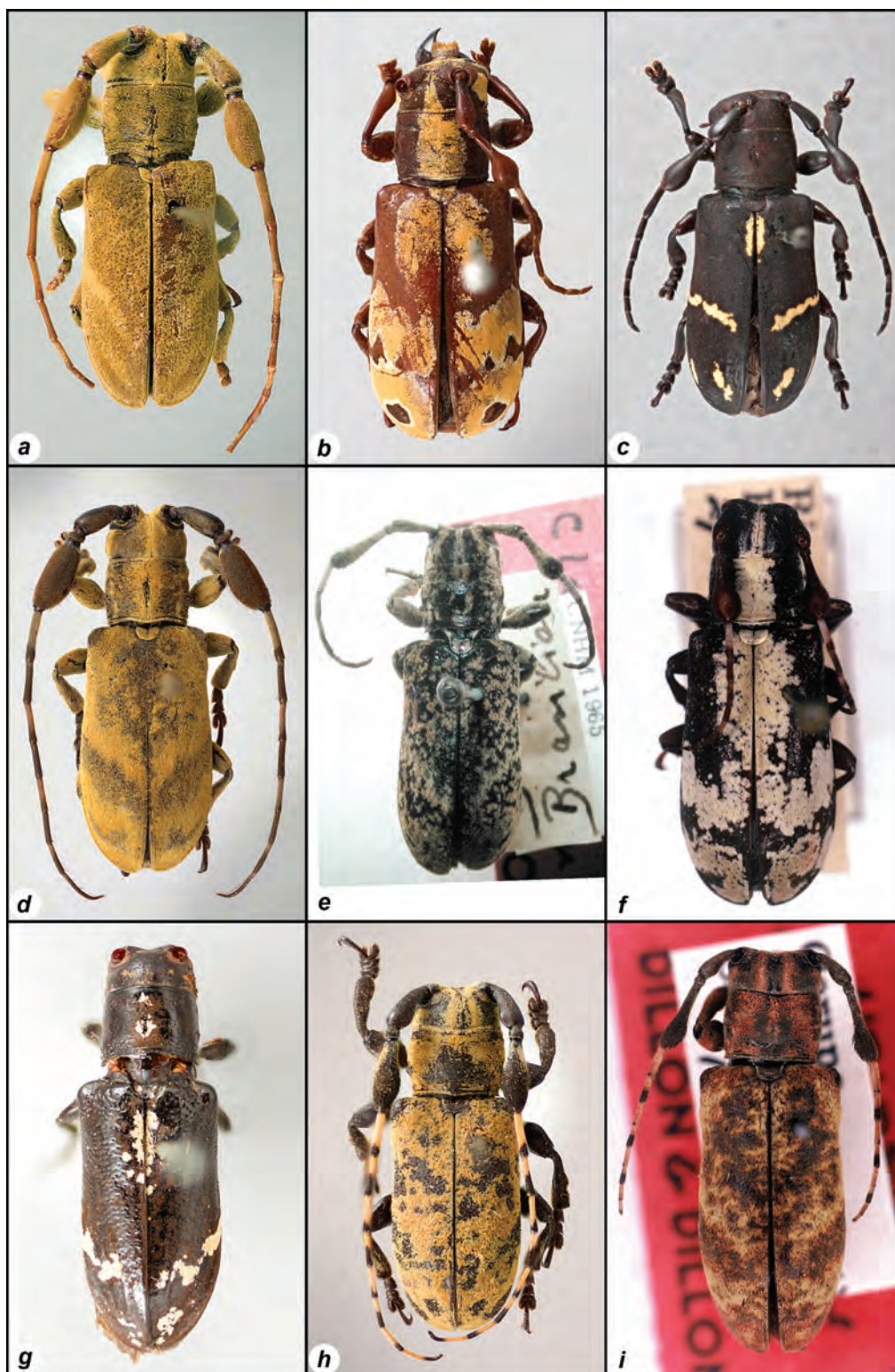


Figure 7. Nine species of *Clavidesmus* Dillon and Dillon, dorsal habitus. **a)** *C. chiccae* Giorgi, 1998, holotype male. **b)** *C. columbianus* Breuning, 1961, holotype female. **c)** *C. funerarius* (Lane, 1958), holotype female. **d)** *C. heterocerus* (Buquet, 1852), holotype male. **e)** *C. indistinctus* Dillon and Dillon, 1952, holotype female. **f)** *C. lichenigerus* (Lane, 1958), holotype female. **g)** *C. metallicus* (Thomson, 1868), holotype female. **h)** *C. monnei* Giorgi, 1998, holotype male. **i)** *C. rubigineus* Dillon and Dillon, 1949, holotype female.

three type specimens of the genus *Peritrox*. Based on examination of the holotype specimen of the type species of the genus (*Peritrox denticollis* Bates, 1865; deposited at the MNHN); and photographs of the type specimens of *Peritrox nigromaculata* Aurivillius, 1920; *Peritrox perbra* Dillon and Dillon, 1945; and *Peritrox vermiculatus* Dillon and Dillon, 1945; the genus *Peritrox* possesses the following characters: eyes large, oblong; frons elongate, about as wide as 1 1/2 lower eye lobe widths; genae distinctly shorter than lower eye lobes, about 1/3 as tall; antennal tubercles armed with short, blunt horn; antennal tubercles narrowly separated, contiguous at base; scape gradually expanded to apex (e.g. Fig. 2a-d).

The three views of the holotype specimen (dorsal habitus, lateral habitus, and close-up of head) provided by Rodrigues and Mermudes (2011) indicate the eyes of *P. insulatus* are moderately large; frons approximately subquadrate, about as wide as 2 1/2 lower eye lobe widths; genae a little shorter than lower eye lobes, about 3/4 as tall; antennal tubercles unarmed, moderately separated, not contiguous at base; scape clavate, not gradually expanded to apex.

Based on these morphological differences, the specimen described by Rodrigues and Mermudes (2011) conforms more closely to characteristics of the genus *Ischiocentra* Thomson, known from Brazil, Colombia, Costa Rica, Panama, and Venezuela. Therefore, we propose the **new combination** *Ischiocentra insulata* (Rodrigues and Mermudes, 2011).

***Jamesia* Jekel, 1861: 259 (Lamiinae: Onciderini)**

(Figures 8a–d)

Jamesia Jekel, 1861: 259. Type species: *Lamia globifera* Fabricius, 1801.

= *Paraclytemnestra* Breuning, 1974: 239. Type species: *Paraclytemnestra gigantea* Breuning, 1974, by monotypy and original designation, **new synonymy**.

Discussion. Fisher (1926) described *Jamesia lineata* (Fig. 8a, c) from a single female specimen from St. Lucia, Lesser Antilles (holotype deposited at USNM). Breuning (1974) described *Paraclytemnestra gigantea* from a single female specimen from the Antilles. Tavakilian (1997) studied both holotype specimens, synonymized *P. gigantea* and proposed the new combination *Paraclytemnestra lineata* (Fisher, 1926). Reexamination of both holotype specimens, as well as dozens of specimens of all described species of *Jamesia*, revealed that both genera share several diagnostic morphological characters: frons elongate, narrow, about as wide as width of one lower eye lobe; frons surface roughly sculptured; eyes with lower lobes oblong; genae about as tall or slightly shorter than lower eye lobes; antennal tubercles narrowly separated, contiguous at base; scape gradually expanded to apex; pronotum transverse, roughly conical, narrower anteriorly (e.g., Fig. 1a-d, 8b, d). Based on these morphological similarities, we propose that *Jamesia lineata* Fisher, 1926 be **restored to original combination**, and *Paraclytemnestra* Breuning becomes a **new synonym** of *Jamesia* Jekel.

***Malthonea* Thomson, 1864: 329 (Lamiinae: Desmiphorini)**

(Figures 9a–d)

Malthonea Thomson, 1864: 329. Type species: *Malthonea tigrinata* Thomson, 1864.

= *Calliphenges* Waterhouse, 1880: 296 (Colobotheni). Type species: *Calliphenges cuprascens* Waterhouse, 1880, by monotypy, **new synonymy**.

Discussion. The monotypic genus *Calliphenges* was described by Waterhouse (1880) and placed in Colobotheni. Examination of the holotype specimen of *Calliphenges cuprascens* Waterhouse, 1880 (collected at Chiguinda, Ecuador and deposited at the BMNH) (Fig. 9a-d) revealed no characters to distinguish it from the genus *Malthonea*. Martins and Galileo (1995) reviewed the genus *Malthonea*, providing diagnostic morphological characters and a key to species. *Calliphenges cuprascens* is here transferred to *Malthonea*, creating the **new combination** *Malthonea cuprascens* (Waterhouse, 1880), and *Calliphenges* Waterhouse becomes a **new synonym** of *Malthonea* Thomson.

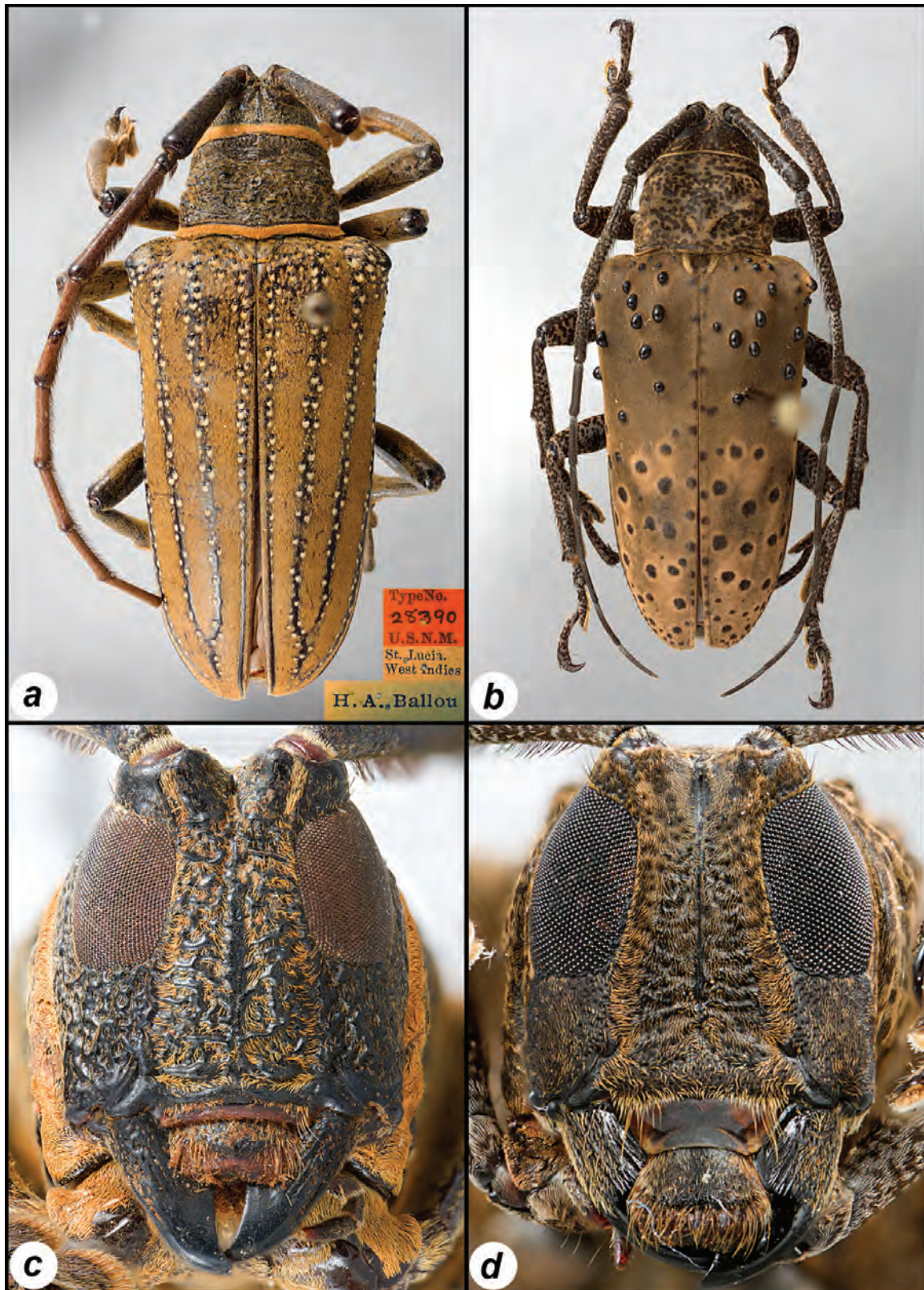


Figure 8. Two species of *Jamesia* Jekel, 1861. **a)** *J. lineata* Fisher, 1926, holotype female, dorsal habitus with original labels. **b)** *J. globifera* Jekel, 1861, female, dorsal habitus. **c)** *J. lineata* Fisher, 1926, holotype, close-up of head. **d)** *J. globifera* Jekel, 1861, female, close-up of head.

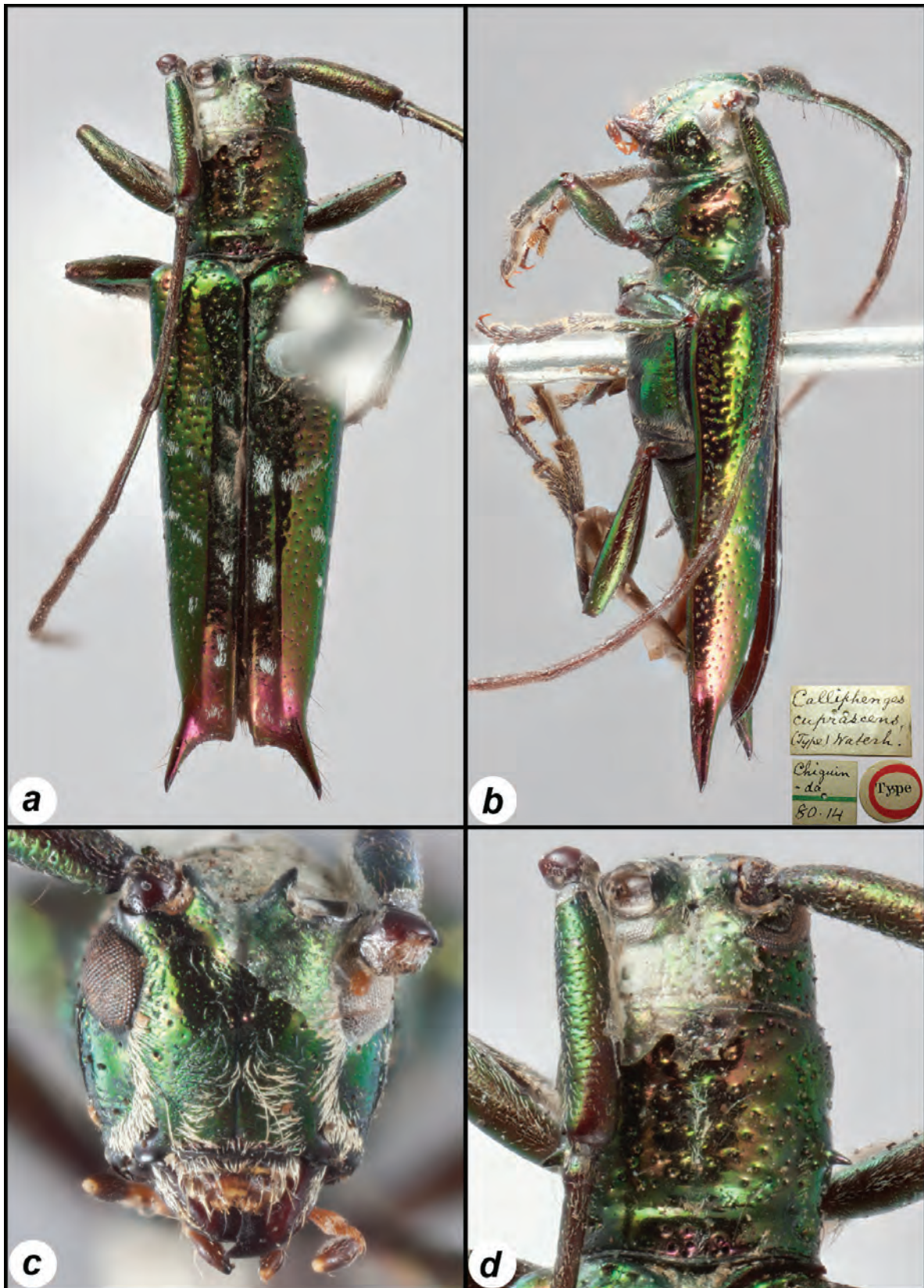


Figure 9. *Malthonea cuprascens* (Waterhouse, 1880), holotype. **a)** Dorsal habitus. **b)** Lateral habitus with original labels. **c)** Close-up of head. **d)** Close-up of pronotum

New Distribution Records in Cerambycidae

Ataxia hovorei Lingafelter and Nearn, 2007 (**Lamiinae: Pteropliini**) is recorded from Haiti, **new country record**. One male specimen (MNHN), "Haïti, ex. museo E. Sallé, 1897, coll. A. Sallé, Museum-Paris." This species was previously believed to be endemic to Dominican Republic (Lingafelter and Nearn 2007; Monné and Bezark 2011).

Carterica soror Belon, 1896 (**Lamiinae: Colobotheini**) is recorded from Ecuador, **new country record**. One specimen (BMNH), "Ecuador: Pichincha, Nambillo Valley near Mindo, 1450m., 13.viii.1987, M. Cooper." This species was previously recorded from Bolivia and Peru (Monné 2005b; Monné and Bezark 2011).

Colobothea lunulata Lucas, 1859 (**Lamiinae: Colobotheini**) is recorded from Colombia, **new country record**. One female specimen (BMNH), "Colombia: Meta, La Macarena, 15.3.76, M. Cooper." This species was previously recorded from Bolivia, Brazil, Ecuador, and Peru (Monné 2005b; Monné and Bezark 2011).

Curius punctatus (Fisher, 1932) (**Cerambycinae: Curiini**) is recorded from Haiti, **new country record**. One female specimen (MNHN), "Haiti, Gonaïves." This species was previously believed to be endemic to Cuba (Monné 2005a; Monné and Bezark 2011; Nearn and Branham 2008).

Cycloplus lacordairei Thomson, 1868 (**Lamiinae: Anisocerini**) is recorded from Colombia, **new country record**. One female specimen (BMNH), "Colombia: Amazonas, Tarapaca, 1/10/76, M. Cooper." This species was previously recorded from Bolivia, Ecuador, French Guiana, and Peru (Monné 2005b; Monné and Bezark 2011).

Iarucanga mimica (Bates, 1866) (**Lamiinae: Hemilophini**) is recorded from Ecuador, **new country record**. One specimen (BMNH), "Ecuador: Morona-Santiago, Cord. de Cutucu 6K. e. of Macas, 1,100m, 12.v.1981, M. Cooper." This species was previously recorded from Bolivia, Brazil, and French Guiana (Monné 2005b; Monné and Bezark 2011).

Pirangoclytus latithorax (Martins and Galileo, 2008) (**Cerambycinae: Clytini**) is recorded from Costa Rica, **new country record**. One specimen (BMNH) "Costa Rica: Guanacaste, Golfito, iii, M. Cooper." This species was previously believed to be endemic to Panama (Monné and Bezark 2011).

Porangonycha princeps (Bates, 1872) (**Lamiinae: Hemilophini**) is recorded from Colombia, **new country record**. One specimen (BMNH), "Colombia: Nariño, Barbarcoas, 40m, 7.X.1990, M. Cooper." This species was previously recorded from Bolivia, Costa Rica, Ecuador, French Guiana, Nicaragua, and Peru (Monné and Bezark 2011; Monné et al. 2012).

Trestonia lateapicata Martins and Galileo, 2010 (**Lamiinae: Onciderini**) is recorded from Brazil, **new country record**. One male specimen (MNHN), "Brésil, Ét. de Goyaz, Jatahy, Museum Paris, Coll. E. Gounelle 1915." This species was previously known from Bolivia (Monné and Bezark 2011).

Tulcus dimidiatus (Bates, 1865) (**Lamiinae: Onciderini**) is recorded from Colombia, **new country record**. One female specimen (MNHN), "S.E. Columbia, Putumayo Superior, Muséum Paris 1932 Col. R. Oberthur." This species was previously known from Brazil and Ecuador (Monné 2005b; Monné and Bezark 2011).

Unaporanga cincta Martins and Galileo, 2007 (**Lamiinae: Hemilophini**) is recorded from Colombia, **new country record**. One male specimen (BMNH), "Colombia: Nariño, Barbarcoas, 40m, 20.iv.1993, M. Cooper." This species was previously recorded from Panama (Monné and Bezark 2011).

Zeale dubia Galileo and Martins, 1997 (Lamiinae: Hemilophini) is recorded from Colombia, **new country record**. One specimen (BMNH), “Colombia: Putamayo, Mocoa, 580m. 5.4.76, M. Cooper.” This species was previously recorded from Bolivia (Monné 2005b; Monné and Bezark 2011).

Zonotylus interruptus (Olivier, 1790) (Cerambycinae: Trachyderini) is recorded from Colombia, **new country record**. One specimen (BMNH), “Colombia: Amazonas, Leticia, 24.8.74, M. Cooper.” This species was previously recorded from Brazil and French Guiana (Monné 2005a; Monné and Bezark 2011; Morvan and Morati 2011: 39, fig. 108).

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Literature Cited

- Bates, H. W. 1865.** Contributions to an insect fauna of the Amazon Valley. Coleoptera: Longicornes. The Annals and Magazine of Natural History 3(15): 213–225, 382–394; 3(16): 101–113, 167–182, 308–314.
- Breuning, S. 1961.** Catalogue des Lamiaires du Monde (Col., Cerambycidae). 4 Lief. Museum G. Frey, Tutzing bei München, p. 183–284.
- Breuning, S. 1974.** Neue Lamiinae (Coleoptera, Cerambycidae). Mitteilungen aus dem Zoologischen Museum in Berlin 50(2): 237–241.
- Dillon, L. S., and E. S. Dillon. 1945.** The tribe Onciderini (Coleoptera: Cerambycidae) Part I. Reading, Scientific Publications, Reading Public Museum and Art Gallery, Number 5: 1–186.
- Dillon, L. S., and E. E. Dillon. 1946.** The tribe Onciderini (Coleoptera: Cerambycidae) Part II. Reading, Scientific Publications, Reading Public Museum and Art Gallery, Number 6: 189–413.
- Fisher, W. S. 1926.** Descriptions of new West Indian longicorn beetles of the subfamily Lamiinae. Proceedings of the United States National Museum 68(22): 1–40.
- Jekel, H. 1861.** Observations suggérées par les notes de M. Chevrolat sur les cérambycides de M. Thomson. Journal of Entomology 1: 255–262.
- Lacordaire 1872.** Histoire Naturelle des Insectes. Genera des Coléoptères, ou exposé méthodique et critique de tous les genres proposés jusqu’ici dans cet ordre d’insectes. Librairie Encyclopédique de Roret, Paris. 9(2): 411–930.

- Lane, F. 1958.** A New Neotropical Genus of Ischiolonchini (Coleoptera, Lamiidae). *American Museum Novitates* 1889: 1–17.
- Lingafelter, S. W., and E. H. Nearns. 2007.** Five new species of longhorned beetles (Coleoptera: Cerambycidae) from the Dominican Republic in genera *Ataxia* Haldeman, *Atimiola* Bates, *Drycothaea* Thomson, *Eburia* Lepeletier and Audinet–Serville, and *Hormathus* Gahan. *The Coleopterists Bulletin* 61(2): 177–191.
- Martins, U. R., and M. H. M. Galileo. 1990.** Onciderini (Coleoptera, Cerambycidae, Lamiinae): sinonimias, novos taxons, claves e notas. *Papeis Avulsos de Zoologia* 37(4): 53–95.
- Martins, U. R., and M. H. M. Galileo. 1995.** Revisão do gênero *Malthonea* Thomson, 1864 (Coleoptera, Cerambycidae, Lamiinae, Desmiphorini). *Revista Brasileira de Entomologia* 39(3): 611–621.
- Monné, M. A. 2005a.** Catalogue of the Cerambycidae (Coleoptera) of the Neotropical Region. Part I. Subfamily Cerambycinae. *Zootaxa* 946: 1–765.
- Monné, M. A. 2005b.** Catalogue of the Cerambycidae (Coleoptera) of the Neotropical Region. Part II. Subfamily Lamiinae. *Zootaxa* 1023: 1–760.
- Monné, M. A. 2012.** Catalogue of the type-species of the genera of the Cerambycidae, Disteniidae, Oxypeltidae and Vesperidae (Coleoptera) of the Neotropical Region. *Zootaxa* 3213: 1–183.
- Monné, M. A., and L. G. Bezark. 2011.** Electronic checklist of the Cerambycidae (Coleoptera) of the Western Hemisphere. Available from: <http://plant.cdfa.ca.gov/bycidb/> (Accessed on 2/2/2012).
- Monné, M. A., E. H. Nearns, S. C. Carbonel Carril, I. P. Swift, and M. L. Monné. 2012.** Preliminary checklist of the Cerambycidae, Disteniidae, and Vesperidae (Coleoptera) of Peru. *Insecta Mundi* 0213: 1–48.
- Morvan, O., and J. Morati. 2011.** Inventaire des Cerambycidae (Coleoptera) de Guyane. I. Parandriinae, Prioninae, Lepturinae et Cerambycinae. In *Contribution à l'étude des Coléoptères de Guyane* 3 (supplément au Bulletin de liaison de l'ACOREP): 10–45, 108 figs, 1 carte.
- Nearns, E. H., and M. A. Branham. 2008.** Revision and phylogeny of the tribes Curini LeConte and Plectromerini Nearns & Branham, new tribe (Coleoptera: Cerambycidae: Cerambycinae). *Memoirs of the American Entomological Society* 47: 1–117.
- Nearns, E. H., and I. P. Swift. 2011.** New taxa and combinations in Onciderini Thomson, 1860 (Coleoptera: Cerambycidae: Lamiinae). *Insecta Mundi* 0192: 1–27.
- Nearns, E. H., N. P. Lord, and K. B. Miller. 2011.** Oncid ID: Tool for diagnosing adult twig girdlers (Cerambycidae: Lamiinae: Onciderini). The University of New Mexico and Center for Plant Health Science and Technology, USDA, APHIS, PPQ. Available from: <http://itp.lucidcentral.org/id/wbb/OncidID/> (Accessed on 2/5/2012).
- Rodrigues, J. M. D. S., and J. R. M. Mermudes. 2011.** A new species of *Peritrox* Bates, with a revised key for the genus (Coleoptera, Cerambycidae, Lamiinae). *Zootaxa* 3000: 44–48.
- Swift, I. P., L. G. Bezark, E. H. Nearns, A. Solís, and F. T. Hovore. 2010.** Checklist of the Cerambycidae and Disteniidae (Coleoptera) of Costa Rica. *Insecta Mundi* 0131: 1–68.
- Tavakilian, G. L. 1997.** Nomenclatural changes, reinstatements, new combinations, and new synonymies among American cerambycids (Coleoptera). *Insecta Mundi* 11(2): 129–139.
- Thomson, J. 1861.** Essai d'une classification de la famille des cérambycides et matériaux pour servir à une monographie de cette famille. Bouchard-Huzard; Paris. 404 p., 3 pls.
- Thomson, J. 1864.** Systema cerambycidarum ou exposé de tous les genres compris dans la famille des cérambycides et familles limitrophes. *Mémoires de la Société Royale des Sciences de Liège* 19: 1–540.
- Waterhouse, C.O. 1880.** New South-American Coleoptera, chiefly from Ecuador. *The Annals and Magazine of Natural History* (5)5: 285–302.

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