LepIntercept

An identification resource for intercepted Lepidoptera larvae

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References

NOCTUIDAE - Spodoptera exigua (Hübner) *Non-Rep*

Taxonomy

Noctuoidea: Noctuidae: Noctuinae: Spodoptera exigua (Hübner)

Common names: beet armyworm, small mottled willow Synonyms: *Caradrina venosa, Laphygma exigua*

Larval diagnosis (Summary)

- Characteristic body coloration (see Figs. 1-7 and Detailed Information)
- Lateral black spot near the SD1 seta on the mesothorax
- Minute sclerotized bar connecting the SD1 setal base to a tonofibrillary platelet on the mesoand metathorax
- Cuticle texture is smooth (at 20X)

Host/origin information

Spodoptera exigua is frequently intercepted from various countries throughout the world, although Mexico is the most common origin, accounting for 75% of the total number of interceptions. Larvae are polyphagous, and approximately 250 hosts are listed in PestID. The most common origin/host combinations are listed here:

Origin	Host(s)
Dominican Republic	Capsicum
Hawaii	Ocimum
Israel	Asclepias, Gerbera
Mexico	Apium, Aster, Brassica, Capsicum, Dianthus, Gladiolus, Helianthus, Mentha, Ocimum, Origanum, Portulaca, Thymus
Netherlands	Asclepias
Thailand	Dendrobium, Oncidium

Recorded distribution

A native of Asia, *Spodoptera exigua* has spread worldwide. It is currently found on every continent except Antarctica, although it do not overwinter in far northern regions and it is rare or absent in parts of South America (Pogue 2002).

Identification authority (Summary)

Being highly polyphagous and cosmopolitan, host/origin data does not help identify *S. exigua* most of the time. Larvae are not expected to be associated with dead plant material or woody conifers. Otherwise, most countries and green plants are potential pathways.

Pest characterization

(Based on Cavey 2001, Pogue 2002)

- Taxonomy: **High.** Species identification is often possible.
- Distribution: Low. Spodoptera exigua is present in the U.S.
- Potential Impact: High. Spodoptera exigua is a pest species.

This ranking characterizes $S.\ exigua$ as not quarantine significant for the U.S.

Larval diagnosis (Detailed)

The larva of the beet armyworm, *Spodoptera exigua*, was partially described by Crumb (1956: 224), Okumura (1961), Peterson (1962), Poque (2002), and Beck (1999-2000). The mouthparts



Fig. 1: Late instar, lateral view



Fig. 2: Late instar, lateral view



Fig. 3: Late instar, lateral view



Fig. 4: Late instar, lateral view



Fig. 5: Mid-instar, lateral view



Fig. 6: Late instar, thorax



Fig. 7: Late instar, thorax

were illustrated by Ahola and Silvonen (2005: 510). Dong et al. (1980) photographed the head and thorax of the first instar.

Typical New World interceptions of *S. exigua* are immediately recognized by the characteristic coloration, lateral black spot near the SD1 seta on the mesothorax and a minute sclerotized bar connecting the SD1 setal base to an adjacent ventral muscle attachment (tonofibrillary platelet) on the meso- and metathorax (Weisman 1974, 1986; Passoa 1991). The cuticle texture is smooth (at 20X).

There are two main color forms. The first one has a darkly marked dorsum with either dashes or rectangular bars, but never with a series of triangular markings (Levy and Habeck 1976, Passoa 1991, Beck 1999-2000: B668, Wagner et al. 2011). The second, more common form, varies from light to dark olive green and is marked with a mixture of spots and dashes on the dorsum (Passoa 1991, Wagner et al. 2011). There is often a large contrast between the pale dorsum and darker subdorsal area (Weisman 1986, Beck 1999-2000: B668).

Several issues complicated identification of *S. exigua* in the New World. Larvae resembling *S. exigua*, but without the mesothoracic black spot, are sometimes intercepted. It is unclear if these are *S. exigua* or not. In other cases, specimens are seen with long pointed spinnerets; these cannot be *S. exigua*. Both of these situations are best identified as "sp. of Noctuidae".

Spodoptera praefica has a rectangular, somewhat obscure, black spot on the mesothorax. This is a western United States species with coloration totally unlike *S. exigua* (see Pogue 2002: 185). Some green forms of *S. frugiperda* resemble *S. exigua* if the pinacula are very pale (Passoa 1991). Crumb (1956) separated these two species, in part, by the position of a line connecting the P1 setae. The line is above the apex if the front in *S. frugiperda* but below the apex in *S. exigua* (see illustrations in Okumura 1961).

More serious is the fact that *S. exigua* is easily confused with early instars of *Copitarsia decolora* and possibly other species of *Copitarsia* as well. The fastest way to seperate these two taxa is by counting the number of thoracic SD sclerotized bars. Spodoptera exigua has only one bar near SD1 whereas *Copitarsia* has two bars, one on SD1 and one on SD2 (Weisman 1986). In addition, the spinneret and labial palpi are different (see data sheet on *Copitarsia* for details).

Weisman (1974) grouped *S. exigua* with two other Old World relatives, *S. exempta* and *S. mauritia*, because all three species have a mesothoracic spot near SD1, no spot on A1 and muscle attachments between the prolegs on A3-6 that form a "Y". The mesothoracic spot on *S. exempta* appears to be absent or obscure at least in some color forms (SPIC). *Spodoptera mauritia* does have a small light brown mesothoracic marking near SD1 (Beardsley 1982). Fortunately, these three species have completely different color patterns and should not be confused with each other. However, it shows that one cannot focus just on a mesothoracic spot to identify *S. exigua* from the Old World.

Identification authority (Detailed)

Being highly polyphagous and cosmopolitan, host/origin data usually does not help identify *S. exigua*. Larvae are not expected to be associated with dead plant material or woody conifers. Most countries and green plants are potential pathways, although in South America it is only reported from Bolivia, Brazil, Peru, and Chile (Zheng et al. 2011).

Specimens preserved in alcohol sometimes lose their color and are very pale. If a lateral mesothoracic spot is discernible, a few backup characters will help confirm *S. exigua*. These include presence of a minute sclerotized bar connecting the SD1 setal base to a ventral muscle attachment on the meso- and metathorax, a bisetose SV group on A1, a smooth cuticle (at 20X) and the muscle attachments between the prolegs on A3-6 forming a short stemmed Y (Passoa 1991). The mandible has no inner teeth or retinaculum. However, none of the morphological characters that define *S. exigua* are very distinctive, and all expected to occur in other noctuid larvae, so specimens with atypical coloration from hosts other than economically important plants need to be examined very carefully. For example, Crumb (1956: couplets 1, 6) listed a few "Amphipyrinae group 7" and "Hadeninae with an open apical silk pore [of the spinneret]" as two taxa with a sclerotized bar on SD1 of the mesothorax. Early instars of *S. exigua* often show the black mesothoracic spot. If other characters fit, these can be identified to species.



Key to recognizing $Spodoptera\ litura/littoralis$ intercepted at U.S. ports of entry (includes $S.\ exigua$)

Origin records

Spodoptera exigua has been intercepted from the following locations:

Argentina, Brazil, Canada, Chile, China, Colombia, Costa Rica, Dominica, Dominican Republic, Ecuador, Egypt, El Salvador, France, Guadeloupe, Guatemala, Haiti, Hawaii, Honduras, India, Israel, Italy, Jamaica, Japan, Kenya, Lebanon, Malaysia, Mexico, Micronesia, Netherlands, Nigeria, Palestinian Territory, Peru, Saudi Arabia, Singapore, South Korea, Spain, St. Lucia, St. Vincent and the Grenadines, Tanzania, Thailand, Trinidad and Tobago, Turkey

Host records

Spodoptera exigua has been intercepted on the following hosts:

Achillea sp., Aconitum sp., Agapanthus sp., Alchemilla sp., Allium ascalonicum, Allium cepa, Allium fistulosum, Allium porrum, Allium sativum, Allium schoenoprasum, Allium sp., Allium tuberosum, Alocasia sp., Alstroemeria aurantiaca, Alstroemeria sp., Amaranthus caudatus, Amaranthus hybridus, Amaranthus sp., Amaranthus spinosus, Ammi majus, Ananas comosus, Anemone sp., Anethum graveolens, Anethum sp., Anigozanthus sp., Antirrhinum majus, Antirrhinum sp.,

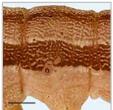




Fig. 8: Abdomen

Fig. 9: Smooth cuticle





Fig. 10: Crochets

Fig. 11: Head





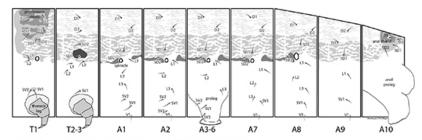
Fig. 12: Hypo. complex

Fig. 13: Mandible

Apiaceae, Apium graveolens, Apium graveolens var. dulce, Apium sp., Aranthera sp., Artemisia dracunculus, Artemisia sp., Artemisia tridentata, Asclepias sp., Asclepias tuberosa, Asparagus officinalis, Asparagus sp., Asparagus sprengeri, Aster ericoides, Aster sp., Asteraceae, Astilbe sp., Bacopa sp., Basella sp., Bergera koenigii, Beta vulgaris var. cicla, Beta vulgaris var. vulgaris, Brassica campestris, Brassica chinensis, Brassica juncea, Brassica napus, Brassica oleracea, Brassica oleracea var. acephala, Brassica oleracea var. alboglabra, Brassica oleracea var. botrytis, Brassica oleracea var. capitata, Brassica oleracea var. italica, Brassica pekinensis, Brassica rapa, Brassica rapa ssp. chinensis, Brassica rapa ssp. pekinensis, Brassica rapa var. parachinensis, Brassica sp., Bupleurum sp., Calendula officinalis, Calendula sp., Callistephus chinensis, Callistephus sp., Capsicum annuum, Capsicum sp., Carthamus sp., Carthamus tinctorius, Celosia argentea, Celosia sp., Chamaemelum sp., Chenopodium album, Chenopodium berlandieri ssp nuttalliae, Chenopodium berlandieri ssp. nuttalliae, Chenopodium sp., Chlorophytum sp., Chrysanthemum sp., Cicer arietinum, Cichorium endivia, Cinnamomum verum, Citrus sp. Clematis sp., Codiaeum variegatum, Colocasia esculenta, Coriandrum sativum, Cucurbita maxima, Cucurbita sp., Cyathea sp., Cynara scolymus, Dahlia sp., Daucus sp., Davallia sp., Delphinium sp., Dendrobium sp., Dianthus barbatus, Dianthus caryophyllus, Dianthus sp., Dysphania ambrosioides, Echeveria sp., Echinodorus sp., Eremurus sp., Ericaceae, Eruca sativa, Eruca sp., Eryngium sp., Eucalyptus sp., Euphorbia sp., Eustoma grandiflorum, Eustoma sp., Fragaria ananassa, Fragaria sp., Genista sp., Gerbera sp., Gladiolus sp., Grevillea sp., Gymnocoronis spilanthoides, Gypsophila elegans, Gypsophila sp., Helianthus annuus, Helianthus sp., Helichrysum sp., Heliconia psittacorum, Heliconia sp., Hydrangea sp., Ipomoea batatas, Iris sp., Lactuca sativa, Lactuca sativa var. capitata, Lactuca sativa var. longifolia, Lactuca sp., Lilium sp. Limonium perezii, Limonium sinuatum, Limonium sp., Lippia sp., Lisianthus sp., Luffa acutangula, Luffa sp., Lycopersicon esculentum, Malvaceae, Matthiola incana, Matthiola sp., Mentha arvensis, Mentha longifolia, Mentha officinalis, Mentha piperita, Mentha sp., Mokara sp., Moluccella laevis, Moluccella sp., Momordica balsamina, Momordica charantia, Momordica sp., Musa sp., Nephelium lappaceum, Nephelium sp., Ocimum basilicum, Ocimum sp., Oncidium basilicum, Oncidium sp., Opuntia sp., Orchidaceae, Origanum majorana, Origanum sp., Origanum vulgare, Peperomia sp., Perilla sp., Petroselinum crispum, Petroselinum sp., Phaseolus sp., Phaseolus vulgaris, Philodendron sp., Phlox sp., Physalis philadelphica, Physalis pubescens, Physalis sp., Pisum sativum, Pisum sativum var. macrocarpon, Pisum sp., Pithecellobium dulce, Polianthes sp., Polianthes tuberosa, Porophyllum tagetoides, Portulaca oleracea, Portulaca sp., Psidium sp. Ranunculus sp., Raphanus sativus, Rosa sp., Rosmarinus officinalis, Rubus sp., Rumex acetosa, Ruscus sp., Saccharum officinarum, Salvia officinalis, Salvia sp., Sechium edule, Solanum lycopersicum var lycopersicum, Solanum sp., Solidago sp., Solidaster sp., Spinacia oleracea, Spinacia sp., Suaeda sp., Symphoricarpos sp., Tagetes erecta, Tagetes sp., Thymus caespititius, Thymus citriodorus, Thymus sp., Thymus vulgaris, Trachelium sp., Tulipa sp., Vaccinium angustifolium, Vaccinium corymbosum, Vaccinium sp., Veronica sp., Viburnum sp., Vigna sesquipedalis, Vigna sp., Vigna unguiculata, Xanthosoma hastifolium, Zea mays, Zingiber officinale

Setal map

Spodoptera exigua (Hübner)



Gilligan, T.M. & S.C. Passoa. 2014. Lepintercept, An identification resource for intercepted Lepidoptera larvae. Identification Technology Program (ITP), USDA/APHIS/PPQ/S&T, Fort Collins, CO (accessed at www.lepintercept.org).

Spodoptera exigua setal map



Click here to download a full-size printable PDF of this larval setal map

LepIntercept - An identification resource for intercepted Lepidoptera larvae by Todd M. Gilligan and Steven C. Passoa Identification Technology Program (ITP), Fort Collins, CO. Last updated February 2014.





